

Bioformulations in Pest Control – A Review

Ana Paula Almeida Portela Silva^{1,2}, Roberto Teixeira Alves³,
Elza Aurea Luna Alves Lima⁴ and Vera Lucia de Menezes Lima^{2*}

¹Biotech Biological Control Company, Maceió, Alagoas, Brazil.

²Department of Biochemistry, Federal University of Pernambuco - UFPE, Recife, Pernambuco, Brazil.

³Laboratory of Ecotoxicology, Embrapa Cerrados, Planaltina, Distrito Federal, Brazil.

⁴Department of Mycology, Federal University of Pernambuco - UFPE, Recife, Pernambuco, Brazil.

Authors' contributions

This work was carried out in collaboration between all authors. Authors APAPS and EALAL designed the work and managed the literature searches. Author APAPS organized figures and conducted the search. Authors APAPS and RTA prepared the initial draft. Author VLML managed cross-opinion corrections, reviewed and proof read the manuscript. All authors read and approved the final manuscript

Article Information

DOI: 10.9734/ARRB/2015/12395

Editor(s):

(1) George Perry, Dean and Professor of Biology, University of Texas at San Antonio, USA.

Reviewers:

(1) Anonymous, University of Idaho, Moscow, USA.

(2) Anonymous, Heilongjiang University, China.

(3) Anonymous, National Research Centre, Dokki, Cairo, Egypt.

(4) Anonymous, University of Calicut, Kerala, India.

Complete Peer review History: <http://www.sciencedomain.org/review-history.php?id=795&id=32&aid=7041>

Mini Review Article

Received 29th June 2014
Accepted 26th September 2014
Published 21st November 2014

ABSTRACT

Biotic and abiotic factors limit the action of entomopathogens and interfere for reaching the expected results. Moreover, the requirement of import and export markets for good quality foods with low content of toxic waste has increased. In this sense, new organic products have been developed in order to improve the stability, virulence and efficacy of entomopathogenic agent in the field. The aim of this paper is to report on the use of formulations with entomopathogenic fungi to control pests. About 12 species or varieties of fungi have been used as active ingredients in formulations of mycopesticides. A formulation can be defined as the combination of an active ingredient (such as entomopathogen), an inert carrier and an adjuvant which will improve the performance of the product, and also will be ease for handling and application. The *Metarhizium anisopliae* and *Beauveria bassiana* are the most used fungi in formulations worldwide. The synergistic effect of fungal interactions with the phytosanitary product has attracted the attention of several researchers

*Corresponding author: E-mail: lima.vera.ufpe@gmail.com;

due to their potential to cause high mortality of the target insect, becoming a tool for deployment in integrated pest management.

Keywords: Formulations with entomopathogens; biological Control; *Metarhizium anisopliae*; *Beauveria bassiana*.

1. INTRODUCTION

Over several decades, the widespread use of chemical insecticides to control pests caused side effects such as environmental imbalance, toxic residues in foods, diseases in humans and other animals and the development of resistance mechanisms in insects [1]. A viable alternative to chemical control is the use of natural enemies, a practice known as biological control. This method enables the maintenance of insect populations in balance, limiting their rapid multiplication without causing harm to other organisms [2,3].

The entomopathogenic fungi are widely used in Biological Control because they are the main pathogens of insects, causing more than 80% of their diseases [4]. Under favorable conditions, they can cause outbreaks and enzootic diseases in natural species of Hemiptera, Lepidoptera and Coleoptera. They specialize in penetration via the tegument and may infect different stages of host development [5-7].

In relation to the Integrated pest management, which recommends the combination of different techniques and resources to maintain a population of insect pests below the economic injury level [8,9], several researches have been designed to enhance the action of entomopathogenic formulations, which contribute to the development of stability, virulence and efficacy of the entomopathogenic agent in the field [10,11]. This study aimed to present a review of the use of formulations containing entomopathogenic fungi to control pests.

2. BIOLOGICAL CONTROL WITH ENTOMOPATHOGENIC FUNGI

Among the entomopathogenic fungi, *Metarhizium*, *Beauveria*, *Paecilomyces*, *Lecanicillium*, *Nomurea*, *Aschersonia*, *Hirsutella* and *Entomophthora* are considered the most important [4] genus. Most of the entomopathogenic fungi have been distributed throughout many decades in the Hyphomycetes class. These organisms were called anamorphic (group of fungi that have no phylogenetic relationship with others, being considered as

perfect) [12]. The anamorph has its counterpart in sexual teleomorph, which corresponds to the sexual phase of Ascomycota or more rarely, Basidiomycota. Subsequently, the *Metarhizium* genus was described as the anamorph of Ascomycota *Cordyceps brittlebankisoides* [13], since the anamorph *Metarhizium anisopliae* var. *majus* (= *M. anisopliae*) [12-15] was isolated from the larvae of Coleoptera (Scarabaeidae). Molecular phylogenetic studies have supported this classification and currently the teleomorph of *Metarhizium* was assigned to *Metacordyceps* [16,17].

Among the entomopathogenic fungi, *Metarhizium anisopliae* (Metsch.) Sorokin and *Beauveria bassiana* (Bals.) Vuillemin stand out due to their wide geographic distribution and host range. The most successful biological control program with *M. anisopliae* was from 1970-1991, when the fungus was applied in several acres of sugar cane fields infested by *Mahanarva posticata* Stal (Hemiptera: Cercopidae), sugar cane leafhopper in the northeast of Brazil [4]. Since then, several researches have been conducted proving the pathogenicity of *M. anisopliae* to other species of agricultural importance such as *Coptotermes formosanus* Shiraki (termites) [18], *Schistocerca gregaria* Forck (grasshoppers) [19], *Hylobius abietis* (pine weevil) [20], *Rhipicephalus (Boophilus) microplus* Canestrini (cattle tick) [21] and *Anthonomus grandis* Boheman (boll weevil) [22].

B. bassiana, an anamorph of *Cordyceps* Fr [23], is one of the most established entomopathogenic fungi taxonomic, and it has widespread occurrence in all countries, being more frequent on insects and soil samples, in which it can survive for long time in saprogenesis [24]. The *B. bassiana* fungus has become internationally known by the Soviet product called Boverin, a wettable powder formulation [25] recommended for the control of *Leptinotarsa decemlineata* Say (potato beetle) and other species. Research conducted with different species showed that *B. bassiana* is pathogenic and virulent to various pests and parasites as *Haematobia irritans* L. (horn fly) [26]; *Psoroptes ovis* Hering (rabbit parasite) [27]; *Laniifera cyclades* Druce (cactus pest) [28]; *Tribolium castaneum* Herbst (red flour

beetle) [29]; *Atteva sciodoxa* Meyrick (medicinal plants caterpillar) [30]; *Hyalomma anatolicum* Koch [31] and *Rhipicephalus (Boophilus) microplus* (ticks) [32], and *Zaprinus indianus* [33].

The mechanism of action of entomopathogenic fungi involves several processes until the insect is completely colonized and killed. First, the mechanical force exerted by the pressure of the hyphae, by breaking the membranous or sclerotic areas of the cuticle of the host, followed by the start of the enzymatic process that results from the release of enzymes, especially proteases, chitinases and lipases, which alter the surface tegument, releasing peptides that serve as nutrients for the fungus and facilitate the penetration into the insect [6,34]. These features are unique to fungi and puts them at an advantage compared to other pathogens that depend on the intake of their propagules to initiate infection [35,7]. Thus, the fungi can infect different host development stages including the stages, which are not fed such as eggs and pupae. After infection, the externalization of the fungus appears on the body surface of the parasitized insect, and the morphologic appearance of the colonization depend on the specie of the entomopathogenic fungi, an example is shown in Fig. 1 for larvae of *Diatraea saccharalis*, the sugarcane borer, killed by *M. anisopliae* (Fig. 1a) and *B. bassiana* (Fig. 1b) colonization. The sick insect dies of a set of modifications on the hemocoele, tissues and internal organs. The cycle is completed when sporulation occurs in the body of the parasitized insect allowing horizontal transmission of the pathogen by spreading the propagules among the insect population, as well as to the environment, so that resulting in spreading infection [36,37].

3. FORMULATIONS OF ENTOMOPATHOGENIC FUNGI

The preservation time and viability of conidia are the main obstacles to its use in a large scale. Generally, biotic and abiotic factors (temperature, solar radiation, moisture, predation competition, among others) limit the fungi action on the field and can have a direct effect on the growth, germination and infective potential of entomopathogenic fungi [38-44]. In this sense, several products have been developed in order to increase stability, virulence and efficacy of the entomopathogenic agent (Fig. 2). Among these products, formulations based on emulsifiable adjuvant oil have been widely studied due to the

facility for storage under controlled temperature and relative humidity ($25\pm 1^{\circ}\text{C}$; $70\pm 10\%$), and protection of conidia against UV rays, with consequent increase persistence in the field and ease of implementation [10].

Several aspects should be considered prior to develop a formulation with entomopathogenic fungi. Firstly, it is necessary the addition of certain compounds that improve the performance of the fungus in the field. Secondly, the formulation should be easier to handle and apply; and finally, that allows a longer time of storage under conditions that minimize the cost, and also with the minimal loss of the quality of the product. These components should also increase the persistence of the product, adhesiveness on the insect and attractiveness to the pest [45] (Fig. 2). In addition to the active ingredient (pathogen) and inert/vector, the formulations can contain adjuvant, components that are used to optimize the action of the active ingredient and improve the characteristics of the formulated product (Fig. 2), for example, their ability to spread on hydrophobic surfaces. Among other properties, they have photoprotective, phagostimulant and anti-evaporation functions.

About 12 species or subspecies (varieties) of entomopathogenic fungi have been used as active ingredients in mycopesticides, for the control of insects and mites, such as *M. anisopliae* var. *anisopliae*, *M. anisopliae* var. *acidum*, *M. flavoviride*, *B. bassiana*, *B. brongniartii*, *Verticillium lecanii*, *Paecilomyces fumosoroseus*, *Isaria farinosa*, *Sporothrix insectorum*, *Hirsutella thompsonii*, *Nomuraea rileyi* and *Cladosporium cladosporioides*. Among the most common products developed worldwide containing entomopathogenic fungi as the active ingredients, *M. anisopliae* and *B. bassiana* are in first place, each one representing 33.9% of the total, followed by *Lecanicillium* spp. (9.4%), *Isaria fumosorosea* (5.8%) and *B. brongniartii* (4.1%). In the inventory of products and formulations, around 26% are substrate colonized by the fungi, 20.5% are wettable powders and 15.2% oil dispersions [45,46].

The most used types of formulation containing entomopathogenic fungi are wettable powder, granules, water dispersible granule, bait, sprinkle powder, powder for contact, oil dispersion, suspension concentrate, miscible suspension concentrate in oil, and suspension in ultra-low volume. The formulation of wettable powder type is applied after dilution with water, such as Boveril[®], a commercial product which is used for

control of pests in crucifers [47]. The sprinkle powder is applied by dusting, whilst powder for contact is by direct application on the pest [45,48]. The granules are a kind of solid formulation of uniform size, but cereal grains such as rice are not included in this type of formulation, because they are considered as technical concentrated or non-formulations [49]. On the other hand, water dispersible granule disintegrates in water before application, such as PFR97 TM[®] having *Paecilomyces fumosoroseus* as the active ingredient [50].

The type bait formulation was developed to attract the pest and be consumed by it, as traps Termitrap[®] used for subterranean termites, water based and sugar cane molasses [51]. The oily dispersion contains the active ingredient (entomopathogenic fungus) in surfactant for using after dilution in water [52]. In this type of formulation are included suspensions in oil

emulsion, for example Met52 EC[®], a commercial product (Novozymes Biologicals, Inc., USA) that must be diluted prior to use in the laboratory or in the field, e.g. against the sweet potato beetle *Cylas formicarius* [53]. The suspension concentrate is already an active ingredient in water, and may be diluted further prior to be applied [54]. The miscible suspension concentrate in oil is a suspension containing the active ingredient in a fluid for dilution in organic liquid, in this formulation it is included the Metarril SP Organic[®] produced by Koppert Biological Systems [55]. The suspension in ultra-low volume comes ready to use or may need small dilution, but it requires special sprayers equipment for application [56], in order to avoid blockage in the outlet nozzle of the applicators; most of ultra-low volume sprayers utilize a small electric pump that can be very finely adjusted to vary droplet size and flow rate, so that meet the desired specific spray application.

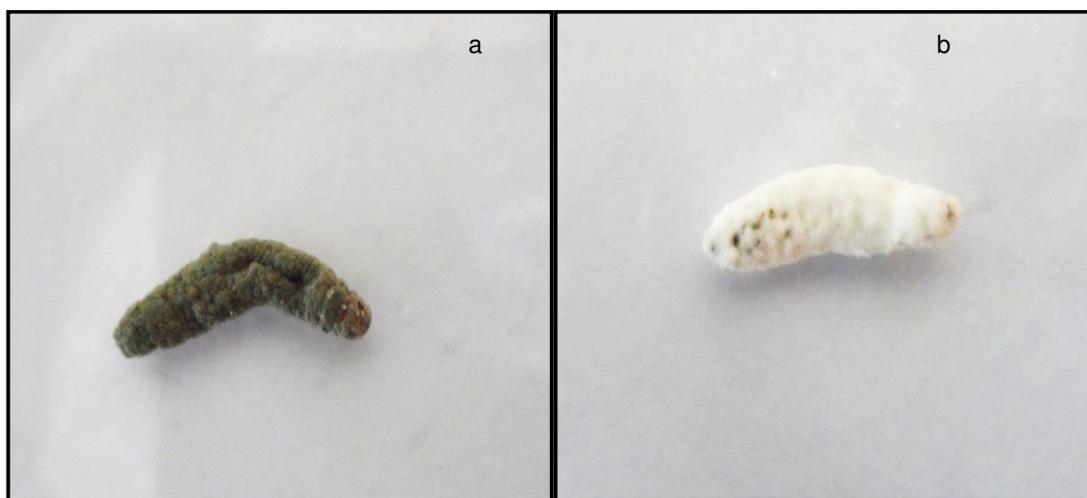


Fig. 1. Larvae of *Diatraea saccharalis* colonized by *Metarhizium anisopliae* (a) and *Beauveria bassiana* (b)

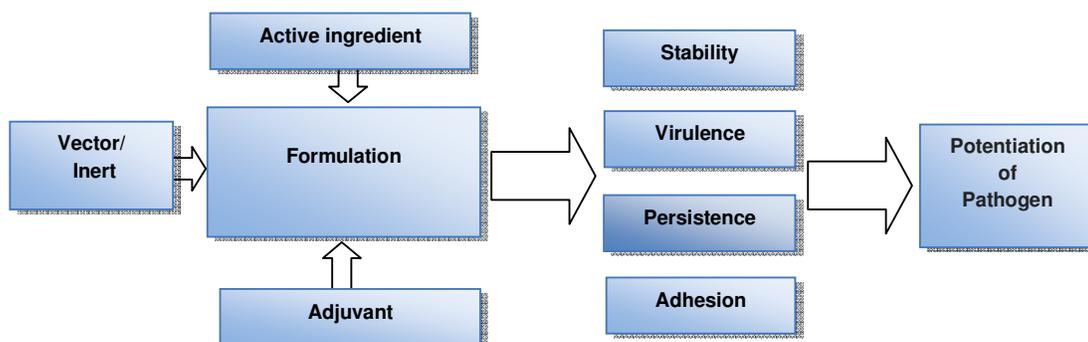


Fig. 2. Schematic representation of development of formulations and their advantages

4. GLOBAL SCENARIO OF FORMULATIONS

In Brazil, products based on *M. anisopliae* represent 55% of commercially available products or in the registration process, followed by *B. bassiana* (30%), *Lecanicillium* spp. (7.5%) and *Sporothrix insectorum* (7.5%). Most of these Brazilian mycopesticides have no record; of these, 2.5% are marketed as pure conidia, 72.5% are technical concentrates (liquid or solid substrates, colonized by fungi) and only 25% are in fact oil dispersion formulations. In this sense, studies with formulations based on vegetable oils and emulsifiers have been carried out in several countries and have produced interesting results for example, when the neem oil (*Azadirachta indica*) is associated with the fungus *B. bassiana*, causing over 90% mortality, on nymphs of *Bemisia tabaci* (whitefly), in the United States of America [57]. Experiments in the South Africa with *Tetranychus urticae* Koch exposed to *B. bassiana* in an oil emulsion obtained 61% of mortality of mites after seven days [58], demonstrating the potential of the formulation for field-testing.

Test formulations of the fungus *M. anisopliae* and *B. bassiana* in oil, for the control of malaria mosquito (*Anopheles* sp.) have showed that formulated conidia were more effective in controlling larvae than non-formulated ones, in addition to persisting longer under field conditions, in the Netherlands [59]. Other studies have also confirmed the high mortality of insects (*Plutella xylostella* L.) when in contact with castor oil (*Ricinus communis* L.) added with *B. bassiana* [60].

Therefore, one can notice that the synergism of the association between fungus and oil, indicate its potential against the target pest in the field. The availability of products on the market formulated with high concentration and viability of infective structures, easy handling and application, greater efficiency and with competitive price is essential for establishing the use of entomopathogenic fungi for pest control in large-scale [61,62].

5. CONCLUSION

Among the main advantages of using bioformulations with entomopathogen fungi for biological control of insect pests we can point out the easiness of production of its infective units on a commercial scale, the simplicity of usage in

field conditions, the low cost of its utilization, and mainly, the reduction on environmental impact [63-67]. Interactions between entomopathogenic fungi with phytosanitary products, such as chemical insecticides (e.g. Decis OC), fungicides (e.g. Manzate 800) or herbicides (e.g. Granoxone) are important to evaluate new formulations, since it can be positive when an additive or synergistic action occurs with the entomopathogen and the product. However, a negative interaction may appear when an antagonistic effect is caused by the inhibition of one of the components, which usually is the active ingredient or entomopathogen. Therefore, prior to consider joint implementation as an effective formulation, there is a need for compatibility testing, seeking more selective products and able to promote the conservation of the pathogen in the field for a longer period of time [68,69].

ACKNOWLEDGEMENTS

The authors of this review article would like to thank *Conselho Nacional de Desenvolvimento Científico e Tecnológico* (CNPq) and the *Coordenação de Aperfeiçoamento de Pessoal de Nível Superior* (CAPES) for helping with the research.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Pourseyed SH, Tavassoli M, Bermousi I, Mardani K. *Metarhizium anisopliae* (Ascomycota: Hypocreales): An effective alternative to chemical acaricides against different developmental stages of fowl tick *Argas persicus* (Acari: Argasidae). *Vet Parasitol.* 2010;172(3-4):305-310.
2. Lv J, Wilson LT, Beuzelin JM, White WH, Reagan TE, Way MO. Impact of *Cotesia flavipes* (Hymenoptera: Braconidae) as an augmentative biocontrol agent for the sugarcane borer (Lepidoptera: Crambidae) on rice. *Biol Control.* 2011;56(2):159-169.
3. Sabbour MM, Abdel-Rahman A. Efficacy of isolated *Nomuraea rileyi* and Spinosad against corn pests under laboratory and field conditions in Egypt. *Ann Rev Res Biol.* 2013;3(4):903-912.
4. Alves SB, Lopes RB, Vieira AS, Tamai MA. Fungos entomopatogênicos usados no

- controle de pragas na América Latina. In: Alves SB, Lopes RB (eds). Controle Microbiano de Pragas na América Latina: avanços e desafios. 1st ed. Piracicaba: FEALQ; 2008.
5. Shubakov A, Kucheryavykh PS. Chitinolytic Activity of Filamentous Fungi. *Appl Biochem Microbiol.* 2004;40(5):445–447.
 6. Donatti AC, Furlaneto-Maia L, Fungaro MHP, Furlaneto MC. Production and Regulation of Cuticle-Degrading Proteases from *Beauveria bassiana* in the Presence of *Rhammatocerus schistocercoides* Cuticle. *Curr Microbiol.* 2008;56(3):256–260.
 7. Kim JS, Roh JY, Choi JY, Wang Y, Shim HJ, Je YH. Correlation of the aphicidal activity of *Beauveria bassiana* SFB-205 supernatant with enzymes. *Fungal Biol.* 2010;114(1):120-128.
 8. Mohan MC, Reddy NP, Devi UK, Kongara R, Sharma HC. Growth and insect assays of *Beauveria bassiana* with neem to test their compatibility and synergism. *Biocontrol Sci Technol.* 2007;17(10):1059–1069.
 9. Lopes RB, Martins I, Souza DA, Faria M. Influence of some parameters on the germination assessment of mycopesticides. *J Invertebr Pathol.* 2013;112(3):236–242.
 10. Alves RT, Bateman RP, Gunn J, Prior C, Leather SR. Effects of different formulations on viability and medium-term storage of *Metarhizium anisopliae*. *Neotrop Entomol.* 2002;31(1):091-099.
 11. Xavier-Santos S, Lopes RB, Faria M. Emulsifiable oils protect *Metarhizium robertsii* and *Metarhizium pingshaense* conidia from imbibitional damage. *Biol Control.* 2011;59(2):261–267.
 12. Kendrick, W.B. The Fifth Kingdom. Canada. Mycologue Publications. 1992;406.
 13. Liu ZY, Liang ZQ, Whalley AJS, Liu AY, Yao YJ. A new species of *Beauveria*, the anamorph of *Cordyceps sobolifera*. *Fungal Divers.* 2001;7:61–70.
 14. Liu Z-Y, Liang Z-Q, Liu A-Y, Yao Y-J, Hyde KD, Yu Z-N. Molecular evidence for teleomorph-anamorph connections in *Cordyceps* based on ITS-5.8S rDNA sequences. *Mycol Res.* 2002;106(9):1100–1108.
 15. Huang B, Li C, Humber RA, Hodge KT, Fan M, Li Z. Molecular evidence for the taxonomic status of *Metarhizium taii* and its teleomorph, *Cordyceps taii* (*Hypocreales*, *Clavicipitaceae*). *Mycotaxon.* 2005;94:137–147.
 16. Sung G-H, Hywel-Jones NL, Sung J-M, Luangsa-ard JJ, Shrestha B, Spatafora JW. Phylogenetic classification of *Cordyceps* and the clavicipitaceous fungi. *Stud Mycol.* 2007;57:5–59.
 17. Bischoff JF, Rehner SA, Humber RA. A multilocus phylogeny of the *Metarhizium anisopliae* lineage. *Mycologia.* 2009;101(4):512–530.
 18. Wright MS, Raina AK, Lax AR. A strain of the fungus *Metarhizium anisopliae* for controlling subterranean termites. *J Econ Entomol.* 2005;98:1451-1458.
 19. Tounou AK, Kooyman C, Douro Kpindou OK, Poehling H.H. Interaction between *Paranosema locustae* and *Metarhizium anisopliae* var. *acidum*, two pathogens of the desert locust, *Schistocerca gregaria* under laboratory conditions. *J Invertebr Pathol.* 2012;97(3):203-210.
 20. Ansari MA, Butt TM. Susceptibility of different developmental stages of large pine weevil *Hylobius abietis* (Coleoptera: Curculionidae) to entomopathogenic fungi and effect of fungal infection to adult weevils by formulation and application methods. *J Invertebr Pathol.* 2012;111(1):33-40.
 21. Quinelato S, Golo PS, Perinotto WMS, Sá FA, Camargo MG, Angelo IC, et al. Virulence potencial of *Metarhizium anisopliae* s.l. isolates on *Rhipicephalus (Boophilus) microplus* larvae. *Vet Parasitol.* 2012;190(3-4):556-565.
 22. Nussenbaum AL, Leucona RE. Selection of *Beauveria bassiana* sensu lato and *Metarhizium anisopliae* sensu lato isolates as microbial control agents against the boll weevil (*Anthonomus grandis*) in Argentina. *J Invertebr Pathol.* 2012;110(1):1-7.
 23. Rehner SA, Minnis AM, Sung GH, Luangsa-Ard JJ, Devotto L, Humber RA. Phylogeny and systematics of the anamorphic, entomopathogenic genus *Beauveria*. *Mycologia.* 2011;103(5):1055-1073.
 24. Sanchez-Peña SR. Entomopathogens from two Chihuahuan desert localities in Mexico. *Biocontrol.* 2000;45(1):63–78.
 25. Ignoffo CM, Garcia C, Alyoshina OA, Lappa NV. Laboratory and Field Studies with Boverin: a Mycoinsecticidal Preparation of *Beauveria bassiana* Produced in the Soviet Union. *J Econ Entomol.* 1979;72(4):562-565.

26. Angel-Sahagún C A, Lezama-Gutiérrez R, Molina-Ochoa J, Galindo-Velasco E, López-Edwards M, Rebolledo-Domínguez O, Cruz-Vázquez C, Reyes-Valázquez WP, Skoda SR, Foster JE. Susceptibility of biological states of the horn fly, *Haematobia irritans*, to entomopathogenic fungi (Hyphomycetes). *J Insect Sci.* 2005;5:1-8.
27. Lekimme M, Focant C, Farnir F, Mignon B, Losson B. Pathogenicity and thermotolerance of entomopathogenic fungi for the control of the scab mite, *Psoroptes ovis*. *Exp Appl Acarol.* 2008;46(1-4):95-104.
28. Lozano-Gutiérrez, España-Luna MP. Pathogenicity of *Beauveria bassiana* (Deuteromycotina: Hyphomycetes) against the White Grub *Laniifera Cyclades* (Lepidoptera: Pyralidae) under Field and Greenhouse Conditions. *Fla Entomol.* 2008;91(4):664-668.
29. Lord JC. Efficacy of *Beauveria bassiana* for control of *Tribolium castaneum* with reduced oxygen and increased carbon dioxide. *J Appl Entomol.* 2009;133(2):101–107.
30. Abood F, Bajwa GA, Ibrahim YB, Sajap AS. Pathogenicity of *Beauveria bassiana* against the Tiger Moth, *Atteva sciodoxa* (Lepidoptera: Yponomeutidae). *J Entomol.* 2010;7(1):19-32.
31. Sun M, Ren Q, Guan G, Liu MA, Gou H, Chen Z, et al. Virulence of *Beauveria bassiana*, *Metarhizium anisopliae* and *Paecilomyces lilacinus* to the engorged female *Hyalomma anatolicum anatolicum* tick (Acari: Ixodidae). *Vet Parasitol.* 2011;180(3-4):389-393.
32. Ren Q, Liu Z, Guan G, Sun M, Ma M, Niu Q et al. Laboratory evaluation of virulence of Chinese *Beauveria bassiana* and *Metarhizium anisopliae* isolates to engorged female *Rhipicephalus (Boophilus) microplus* ticks. *Biol Control.* 2012;63(2):98-101.
33. Svedese VM, Portela-Silva APA, Lopes RS, Santos JF, Luna-Alves Lima EA. Action of entomopathogenic fungi on the larvae and adults of the fig fly *Zaprionus indianus* (Diptera: Drosophilidae). *Cienc Rural.* 2012;42(11):1916-1922.
34. St. Leger RJ, Goettel M, Roberts DW, Staples RC. Penetration events during infection of host cuticle by *Metarhizium anisopliae*. *J Invertebr Pathol.* 1991;58:168-179.
35. Fang W, Feng J, Fan Y, Zhang Y, Bidocha MJ, St Leger RJ, et al. Expressing a fusion protein with protease and chitinase activities increases the virulence of the insect pathogen *Beauveria bassiana*. *J Invertebr Pathol.* 2009;102(1):155–159.
36. Quesada-Moraga E, Martín-Carballo I, Garrido-Jurado I, Santiago-Álvarez C. Horizontal transmission of *Metarhizium anisopliae* among laboratory populations of *Ceratitis capitata* (Wiedemann) (Diptera: Tephritidae). *Biol Control.* 2008;47(1):115–124.
37. García-Munguía AM, Garza-Hernández JA, Rebollar-Tellez EA, Rodríguez-Pérez MA, Villanueva FR. Transmission of *Beauveria bassiana* from male to female *Aedes aegypti* mosquitoes. *Parasite Vector.* 2011;4(24):1-6.
38. Fernandes EK, Rangel DE, Moraes AM, Bittencourt VR, Roberts DW. Variability in tolerance to UV-B radiation among *Beauveria* spp. isolates. *J Invertebr Pathol.* 2007;96(3):237-243.
39. Huang BF, Feng MG. Comparative Tolerances of Various *Beauveria bassiana* isolates to UV-B Irradiation with a Description of a Modeling Method to Assess Lethal Dose. *Mycopathologia.* 2009;168(3):145-152.
40. Bouamama N, Vidal C, Fargues J. Effects of fluctuating moisture and temperature regimes on the persistence of quiescent conidia of *Isaria fumosorosea*. *J Invertebr Pathol.* 2010;105(2):139-144.
41. Ment D, Gindin G, Glazer I, Perl S, Elad D, Samish M. The effect of temperature and relative humidity on the formation of *Metarhizium anisopliae* chlamydospores in tick eggs. *Fungal Biol.* 2010;114(1):49-56.
42. Tiago PV, Carneiro-Leão MP, Malosso E, Oliveira NT, Luna-Alves Lima EA. Persistence and effect of *Metarhizium anisopliae* in the fungal community of sugarcane soil. *BioControl.* 2012;57(5):653–661.
43. Batista-Filho A, Alves SB, Alves LFA, Pereira RM, Augusto NT. Formulação de entomopatógenos. In: S.B. ALVES (ed). *Controle Microbiano de Insetos*. 2nd ed. Piracicaba: FEALQ; 1998.
44. Faria MR, Wraight SP. Mycoinsecticides and mycoacaricides: a comprehensive list with worldwide coverage and international classification of formulation types. *BioControl.* 2007;43(3):237-256.
45. Michereff-Filho M, Faria M, Wraight SP, Silva KFAS. Mycoinsecticides and

- mycoacaricides in Brazil: how are we after four decades? Arq Inst Biol. 2009;76(4):769-779.
46. Etheimine MO, Kane, CMH, Ely SO, Barry A, Mohamed SO, Babah MAO, et al. Storability of five new formulations of Green Muscle® (*Metarhizium acridum*) under ambient and low temperatures: evaluation of conidial viability and virulence against desert locust nymphs. Int J Trop Insect Sci. 2013;33(3):195-201.
 47. Rondelli VM, Pratissoli D, Marques EJ, Júnior HJGS, Alencar JRCC, Sturm GM, Paes JPP. Selection of *Beauveria bassiana* isolates potential for diamondback moth control. Horticultura Brasileira. 2012;30(3):391-396.
 48. Almeida JEM, Rocha TC, Batista-Filho A. Development of a method for physical extraction of conidia of *Metarhizium anisopliae* and *Beauveria bassiana* to dry and wettable powder formulation of biopesticides. Arq Inst Biol. 2007;74(4):369-371.
 49. Freitas AF, Loureiro ES, Almeida MEB, Pessoa LGA. Yield of conidia and germination of different isolates of *Metarhizium anisopliae* (Metsch) Sorok. (Ascomycota: Clavicipitaceae) grown on rice. Arq Inst Biol. 2014;81(1):75-78.
 50. Faria MR, Magalhães BP. O uso de fungos entomopatogênicos no Brasil. Biotecnologia. Cienc. Desenvolv. 2001;22:18-21. Available: <http://www.biotecnologia.com.br/revista/bio22/fungos.pdf>
 51. Almeida JEM, Alves SB, Moino Jr A, Lopes RB. Control of the termite *Heterotermes tenuis* (Hagen) using termitrap baits with insecticides associated with the entomopathogenic fungus *Beauveria bassiana* (Bals.) Vuill. An. Soc. Entomol. 1998;27(4):639-644.
 52. Suyambulingam AK, Subbiah SN, Kannan R, Rajamanickam C, Sengottayan SN. Effect of oil-formulated *Metarhizium anisopliae* and *Beauveria bassiana* against the rice leaf folder *Cnaphalocrocis medinalis* Guenée (Lepidoptera: Pyralidae). Arch Phytopathol PFL. 2014;47(8):977-992. DOI: 10.1080/03235408.2013.828388
 53. Reddy GVP, Zhao Z, Humber RA. Laboratory and field efficacy of entomopathogenic fungi for the management of the sweetpotato weevil *Cylas formicarius* (Coleoptera: Brentidae). J Invertebr Pathol. 2014;122:10-15.
 54. Li MY, Lin HF, Li SG, Xu AM, Feng MF. Efficiency of entomopathogenic fungi in the control of eggs of the brown planthopper *Nilaparvata lugens* Stål (Homoptera: Delphacidae). Afr. J. Microbiol. Res. 2012;6(44):7162-7167. DOI: 10.5897/AJMR12.611.
 55. Camargo MG, Marciano AF, Sá FA, Perinotto WMS, Quinelato S, Gólo OS, Angelo IC. Commercial formulation of *Metarhizium anisopliae* for the control of *Rhizoglyphus microplus* in a pen study. Vet Parasitol. 2014;205(3):271-276.
 56. Alves RT, Bateman RP. Delivery systems for Mycoinsecticides Using Oil-based Formulations. Aspect Appl Biol. 2000;57:163-170.
 57. Islam MT, Castle SJ, Ren S. Compatibility of the insect pathogenic fungus *Beauveria bassiana* with neem against Sweet potato whitefly, *Bemisia tabaci*, on eggplant. Entomol Exp Appl. 2010;134(1):28-34.
 58. Gatarayihya MC, Laing Md, Miller RM. Effects of adjuvant and conidial concentration on the efficacy of *Beauveria bassiana* for the control of the two spotted spider mite, *Tetranychus urticae*. Exp Appl Acarol. 2010;50(3):217-229.
 59. Bukhari T, Takken W, Koenraadt CJM. Development of *Metarhizium anisopliae* and *Beauveria bassiana* formulations for control of malaria mosquito larvae. Parasite Vector. 2011;4(23):1-14. DOI: 10.1186/1756-3305-4-23
 60. Rondelli VM, Pratissoli D, Polanczyk RA, Marques EJ, Sturm GM, Tiburcio MO. Association of castor bean oil with *Beauveria bassiana* for diamondback moth control. Pesq Agropec Bras. 2011;46(2):212-214.
 61. Orlandelli RC, Pamphile JA. Entomopathogenic fungus *Metarhizium anisopliae* as biological control agent of insect-pests. SaBios: Rev Saude e Biol. 2011;6(2):79-82. Available: <http://revista.grupointegrado.br/revista/index.php/sabios2/article/view/757>
 62. Farenhorst M, Knols BG, Thomas MB, Howard AFV, Takken W, Rowland M, Guegan RN. Synergy in efficacy of fungal entomopathogens and permethrin against west african insecticide-resistant *Anopheles gambiae* mosquitoes. Plos One. 2010;5(8):e12081.
 63. Lopes RB, Martins I, Souza DA, Faria M. Influence of some parameters on the germination assessment of

- mycopesticides. J Invertebr Pathol. 2013;112(3):236-242.
64. Nussenbaum AL, Lewylle MA, Lecuona RE. Germination, Radial Growth and Virulence to Boll Weevil of Entomopathogenic Fungi at Different Temperatures. World Appl Sci J. 2013;25(8):1134-1140.
65. Borisade OA, Magan N. Growth and sporulation of entomopathogenic *Beauveria bassiana*, *Metarhizium anisopliae*, *Isaria farinosa* and *Isaria fumosorosea* strains in relation to water activity and temperature interactions. Biocontrol Sci Technol. 2014;24(9):999-1011.
66. Blanford S, Shi W, Christian R, Marden JH, Koekemoer LL, Brooke BD, et al. Lethal and pre-Lethal effects of a fungal biopesticide contribute to substantial and rapid control of malaria Vectors. Plos One. 2011;6(8):e23591.
67. Zahran HEDM, Kawanna MA, Bosly HA. Larvicidal Activity and Joint Action Toxicity of Certain Combating Agents on *Culex pipiens* L. Mosquitoes. Ann Rev Res Biol. 2013;3(4):1055-106.
68. Borges LR, Vila Nova MX. Association of chemical insecticides and entomopathogenic fungi in Integrated Pest Management – a review. Ambiência. 2011;7(1):179–190. Available:<http://revistas.unicentro.br/index.php/ambiencia/article/viewFile/546/1200>.
69. Vidau C, Diogon M, Aufavre J, Fontbonne R, Viguès B, Brunet JL, et al. Exposure to sublethal doses of Fipronil and Thiacloprid highly increases mortality of honeybees previously infected by *Nosema ceranae*. Plos One. 2011;6(6):e21550.

© 2015 Silva et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:
<http://www.sciencedomain.org/review-history.php?iid=795&id=32&aid=7041>