

Anti-bacterial Effect of Aqueous and Ethanolic Extracts of Onion, Garlic and Cinnamon on *Xanthomonas* Species

**Amir Hussain¹, Tahir Naqqash^{2*}, Kashif Aslam², Syed Bilal Hussain²,
Shahid Masood Shah³ and Ghulam Shabir^{2*}**

¹Department of Molecular Biology, Virtual University, Multan, Pakistan.

²Institute of Molecular Biology and Biotechnology, Bahauddin Zakariya University,
60800 Multan, Pakistan.

³Department of Biotechnology, COMSATS University Islamabad, Abbottabad Campus, Pakistan.

Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/BJI/2021/v25i430148

Editor(s):

- (1) Dr. Chan Yean Yean, Universiti Sains Malaysia, Malaysia.
(2) Dr. Pankaj Kumar, H. N. B. Garhwal Central University, India.
(3) Prof. Antar El-Banna, Kafrelsheikh University, Egypt.

Reviewers:

- (1) S. Selvarani, Thiagarajar College, India.
(2) Lalu Zulkifli, University of Mataram, Indonesia.

Complete Peer review History: <https://www.sdiarticle4.com/review-history/74590>

Original Research Article

**Received 09 August 2021
Accepted 18 October 2021
Published 28 October 2021**

ABSTRACT

Onion (*Allium cepa*), garlic (*Allium sativum*) and cinnamon (*Cinnamomum zeylanicum*) are edible herbs but they contain many chemicals which can be used as medicine. These herbs also contain many antibiotic molecules that are capable to inhibit or reduce growth of many types of microbes including bacteria. Due to the development of resistance of bacteria against synthetic antibiotic now there is a need to have natural antibiotic which must be harmless, cheap and easily available. To know the effectiveness of garlic, onion, and cinnamon on disease causing bacteria such as *Xanthomonas campestris* pv. *malvacearum*, *Xanthomonas axonopodis* pv. *citri*, *Xanthomonas oryzae* pv. *oryzae*, extracts of each sample was made in water and ethanol. Antibacterial potential of plant extracts was observed at two different stages: at the time of inoculation and after growth of pathogen on media. Results showed extracts applied on all the bacteria before growth inhibited

*Corresponding author: E-mail: tahirnaqqash@gmail.com, shabirpbg50@gmail.com;

maximum bacterial growth as compared to applied after growth. Moreover, cinnamon extract in ethanol effectively control bacterial growth than onion and garlic extracts. Maximum inhibition zone was shown by ethanolic extract of cinnamon before growth of *Xanthomonas oryzae pv. oryzae* (7.333 mm), *Xanthomonas campestris pv. malvacearum* (7.83 mm) and *Xanthomonas axonopodis pv. citri* (5.1 mm). After 24 hours, maximum growth of *Xanthomonas oryzae* and *Xanthomonas axonopodis* was inhibited by ethanolic extracts of cinnamon (2.82 and 3.35 mm, respectively) while *Xanthomonas campestris* was inhibited by ethanolic extract of onion (6.55 mm). Extract diluted from 66mg/ml to 0.01mg/ml showed different minimum inhibitory concentration against pathogens. As concentration decreases inhibition of bacteria also decreases. Ethanolic extracts of cinnamon showed maximum MIC against *Xanthomonas campestris pv. malvacearum* (3.5 mm to 1.5 mm) and *Xanthomonas oryzae pv. oryzae* (5 mm to 2 mm) while for *Xanthomonas axonopodis pv. citri* ethanolic extract of garlic (3 mm to 1 mm) was effective ranging between 66mg/ml to 33 mg/ml. This study highlighted that natural products possess ability to inhibit pathogenic bacterial growth and would also be helpful in medicinal field for further study.

Keywords: Antibacterial activity; ethanolic extract; aqueous extract; spices.

1. INTRODUCTION

Many important crops are infected by bacteria such as *Xanthomonas* species which damages more than 50% percent production of rice, cotton and citrus etc. [1]. *Xanthomonas* (from the Greek xanthos, meaning, "yellow" and monas, meaning entity) is associated with plant yellow pigment bacteria which belong to large genus of gram-negative bacteria. *Xanthomonas* has the largest species that affect nearly four hundred plants and belong to class of Gamma proteobacteria. *Xanthomonas* damages most important economical crops such as rice, citrus, banana, cabbage, tomato, pepper and bean. It is host specific bacteria and infects plants with contaminated seed, rainwater, or aerosols. Bacteria remain on the surface of leaf or stem then enter into parenchyma cells through stomata, wounds and hydathodes. Approximately all species of *Xanthomonas* contain chromosomes ranging from 4-5.3 mega base pairs with single circular and GC content nearly 70% out of hundred almost all genes are same but in genomic sequences large number of inversions, insertion, translocation, deletion [2].

Xanthomonas axonopodis pv. citri destroys most economical citrus fruits, 125 monocot and 268 dicot plants. This disease known over centuries but disease-causing agent was first noticed in India [3]. Citrus production in many countries of the world decreases because of this bacterial attack. *Xanthomonas axonopodis* or citrus canker develops lesions on all parts of the plant, due to this fruit quality and yield reduced. This disease prevails all over US as well as in Asia due to this [4]. Bacterial leaf blight (*Xanthomonas campestris pv. malvacearum*) affects all aerial

parts of the cotton crop, causes premature defoliation, blockage of xylem phloem tissues and finally death of parenchyma tissues [5]. Pakistan's is the third biggest exporter of cotton and due to this the production affects badly that ultimately affects Pakistan economy [6]. Rice is most abundantly cultivated across the world in temperate regions but Asia produces rice 90% of the world. Pakistan is the second country for the production of rice in world. However, this most important crop is destroyed by bacterial leaf blight disease caused by *Xanthomonas oryzae pv. oryzae* all around the world. This disease may attack at any stage of crop development which results in immature grain and total yield lost [7].

Most of our agricultural industry treats crop diseases with different chemicals that are non-biodegradable and leach down in soil, move into water resources and their residue remains in edible parts of crop. These chemicals add toxic components to environment and are inhaled by human and animals causing different lethal diseases related to breathe, stomach heart and cancer. Contaminated water also causes difficulties for the survival of aquatic animals. In Pakistan, bacterial blight is being treated with strong and environmental toxic chemicals such as Copper- oxychloride which is expensive and its residuals pose danger to human health. Now a day's researchers are giving attention to environmental friendly and nontoxic methods for treating plant diseases. For this purpose, developing countries like Pakistan use plants extract for treating different diseases that do not affect the growth and production of crops. Interestingly about 20% of plants extract contain such metabolites that are used against bacterial diseases [7].

The use of medicinal plants or herbs has received greater attention due to its minimum or no adverse effects. Spices are substances used to increase food taste and aroma including leaves (coriander and mint), bulbs (turmeric and garlic), fruits (black pepper), flowers (clover), rhizomes (ginger) and stems (cinnamon) are some of the plant components utilized to improve taste. Several bioactive compounds produced by medicinal plants have antibacterial and antifungal properties [8]. Plants contain a wide range of secondary metabolites, including tannins, terpenoids, alkaloids, and flavonoids, all of which were shown to have antibacterial effects [7]. Numerous medicinal plants also include antioxidant and antimicrobial characteristics that defend the host against cellular oxidation processes and other infections, emphasizing the significance of exploring the natural antimicrobials [9]. The majority of bacterial infections found in foods are susceptible to plant extracts such as oregano, garlic, mustard, cinnamon and onion [10]. Among these garlic (*Alliums sativum*) is being used as medicine for long time due to its antibiotic, antifungal and antiviral properties [11]. Garlic is used as herbal antioxidant that reduces reactive oxygen species and low-density lipoprotein oxidize it containing allicin and sulfur compound that are highly active against microbes [12,13]. Onion (*Allium cepa*) grows worldwide and has strong antiseptic quality due to different chemicals in its skin and bulb such as quercetin, aglycone, flavonoids, phenols and polyphenols. As an anti-bacterial agent its production has been increased around 25% in last 10 years [14]. Cinnamon (*Cinnamomum zeylanicum*) is a green spice plant that is grown in tropical region and also used as medicine in ancient and modern ages. Bark and leaves contains chemicals like cinnamte, cinnamic acid, and cinnamaldehyde that have anti-diabetic, anti-fungal, antioxidant, anti-inflammatory, anticancer, insecticidal, and nematocidal properties [15]. Based on the above literature, present study is planned to investigate the anti-bacterial activity of onion, garlic and cinnamon extract on *Xanthomonas* species causing damage to economically important crops.

2. MATERIALS AND METHODS

2.1 Sample Collection

Fresh onion, garlic and dry cinnamon were purchased from local market. Onion and garlic were disease free and without physical damage. *Xanthomonas campestris* pv. *malvacearum*,

Xanthomonas oryzae pv. *oryzae*, *Xanthomonas axonopodis* pv. *citri* were collected from Department of Agriculture, Bahauddin Zakariya University, Multan, Pakistan.

2.2 Preparation of Extract

For extract preparation, onion and garlic were peeled, washed and dried. All the clean samples were cut into small pieces and were grounded in mortar and pestle. Dry Cinnamons were grounded in electric grinder and powder was made. 40 g of each sample was taken and soaked in 400 ml of water and 400 ml ethanol in separate beakers. Each sample was left for 72 hours and stirred at 25°C using magnetic stirrer. These water and ethanol extracts were centrifuged at 3000 rpm for 10 minutes at room temperature. Each extract prepared was weighed i.e., water and ethanol extract of cinnamon were 2.015 g and 3.595 g, respectively, water and ethanol extract of onion were 2.033g and 2.022 g, respectively, and water and ethanol extract of garlic was 2.031 g and 2.019 g, respectively.

2.3 Media Preparation and Bacterial Inoculation

Antibacterial potential of plant extracts was tested at two different stages of growth by applying the extract at the time of inoculation and after the growth of pathogen on media. For this purpose, pure culture of *Xanthomonas campestris* pv. *malvacearum*, *Xanthomonas oryzae* pv. *oryzae* and *Xanthomonas axonopodis* pv. *citri* were cultured in nutrient broth under highly sterile conditions following streaking on nutrient agar plates and were placed for 48 hours at 28°C. After that, these cultures were poured on nutrient agar plates and placed in incubator at room temperature for 24hours. All the activities were performed in laminar flow to prevent entrance of irrelevant microbes.

2.4 Extract Application before Bacterial Growth

Nutrient agar plates were taken and bacterial suspension was applied on petri-dishes evenly before placing extracts dip disc placed at different position on petri dishes. Paper discs were dipped in already prepared aqueous and ethanolic extract of garlic, cinnamon, and onion having the concentration of 100 mg/ml for two minutes and were placed in the culture plates for two days. For negative control group, water and ethanol dipped disc were placed on the culture

plates while for positive control penicillin (30µg) was used and results were noticed. The experiment was repeated three times. A precise ruler was used to determine the zone of inhibition as strong (10-20 mm), moderate (5-10 mm) or weak (1-5 mm) based on the diameter of the inhibition zone [16].

2.5 Extract Application after Bacterial Growth

When bacterial growth appeared on culture plates, extract was applied by using disc diffusion method. 8 mm disc was dipped in ethanol and water extract of garlic, cinnamon, and onion having the concentration of 100 mg/ml for 2 minutes and was placed on culture plates of each bacterium. Extracts applied on the bacterial pathogen's samples were placed in incubator for 24 hours. For negative control group, water and ethanol dipped disc were placed on the culture plates while for positive control penicillin (30 µg) was used and results were noticed. The experiment was repeated three times. A precise ruler was used to determine the zone of inhibition as strong (10-20 mm), moderate (5-10 mm) or weak (1-5 mm) based on the diameter of the inhibition zone [16].

2.6 Minimum Inhibitory Concentration (MIC)

The broth dilution technique was used to determine the MIC [17]. The aqueous and ethanolic extracts were diluted in distilled water in different concentrations (66.6 to 0.01 mg/ml). In the test tubes, 500 µl of different concentrations aqueous and ethanolic extracts were introduced in sterile nutritional broth (2 ml) along with the test microorganisms (1 ml) following incubation for 24 hours at 37°C. The minimum inhibitory concentration was determined as the lowest extract concentration that prevented observable growth of each of the test microorganisms.

2.7 Statistical Analysis

All of the tests were repeated three times, with the findings reported as mean ± standard deviation. Data was examined statistically by Statistics 8.1 software using analysis of variance (ANOVA) approach at 5% probability. The least significant difference (LSD) test was used to compare means.

3. RESULTS AND DISCUSSION

Xanthomonas spp. damage most crops and fruits that are economically important. Their control

with chemicals is more expensive which may reduce the quality of natural products likewise [1]. Control of these bacteria is very important for protecting crops and plants. Natural plants (cinnamon, onion, garlic) extracts are also used to control the growth of such bacteria. All extracts used in this study successfully controlled the growth of these bacteria.

3.1 Before Inoculum of Bacteria

Results for test bacterial growth inhibition zone before grow this shown in Fig. 1A to C. The penicillin used as a positive control showed significant activity on all the three isolates as compared to the aqueous and ethanolic extracts of all three spices, however, water and ethanol taken as negative control showed zero activity against isolates. Among all three spices, maximum inhibition zone was shown by ethanolic and aqueous extract of cinnamon against *Xanthomonas oryzae* pv. *oryzae* (7.333 mm and 3.5 mm), *Xanthomonas campestris* pv. *malvaceum* (7.83 mm and 7.33 mm) and *Xanthomonas axonopodis* pv. *citri* (5.1 mm and 2.333 mm) as shown in Fig. 1A. Thus, it can be concluded that cinnamon extract was more effective to control the growth than garlic and onion extracts. In comparison to aqueous extract, the cinnamon ethanolic extract has higher antibacterial activity. The reasoning behind this is that the antibacterial substance of cinnamon bark show more solubility in ethanol than in water as suggested in previous studies [18,19].

3.2 After Growth of Bacteria

When bacteria were grown for 24 hours and covered culture plates were treated with all three extracts mentioned above they showed different zone of inhibition as shown in Fig. 2A to 2C. The penicillin used as a positive control showed significant activity on all the three isolates as compared to the aqueous and ethanolic extracts of all three spices, however, water and ethanol taken as negative control showed zero activity against isolates. Maximum growth of *Xanthomonas oryzae* was inhibited by ethanolic (2.82 mm) and aqueous (1.65 mm) extracts of cinnamon as compared to garlic and onion extracts. However, maximum growth of *Xanthomonas campestris* was inhibited by ethanolic extract of onion (6.55 mm) and aqueous extract of garlic (3.75mm) as compared to other extracts. In the case of *Xanthomonas axonopodis* ethanolic extract of cinnamon (3.35 mm) and aqueous extract of garlic (2.50 mm)

showed maximum growth inhibition. These results are similar to previous findings conducted in different bacterial species such as *E.coli* [19,20].

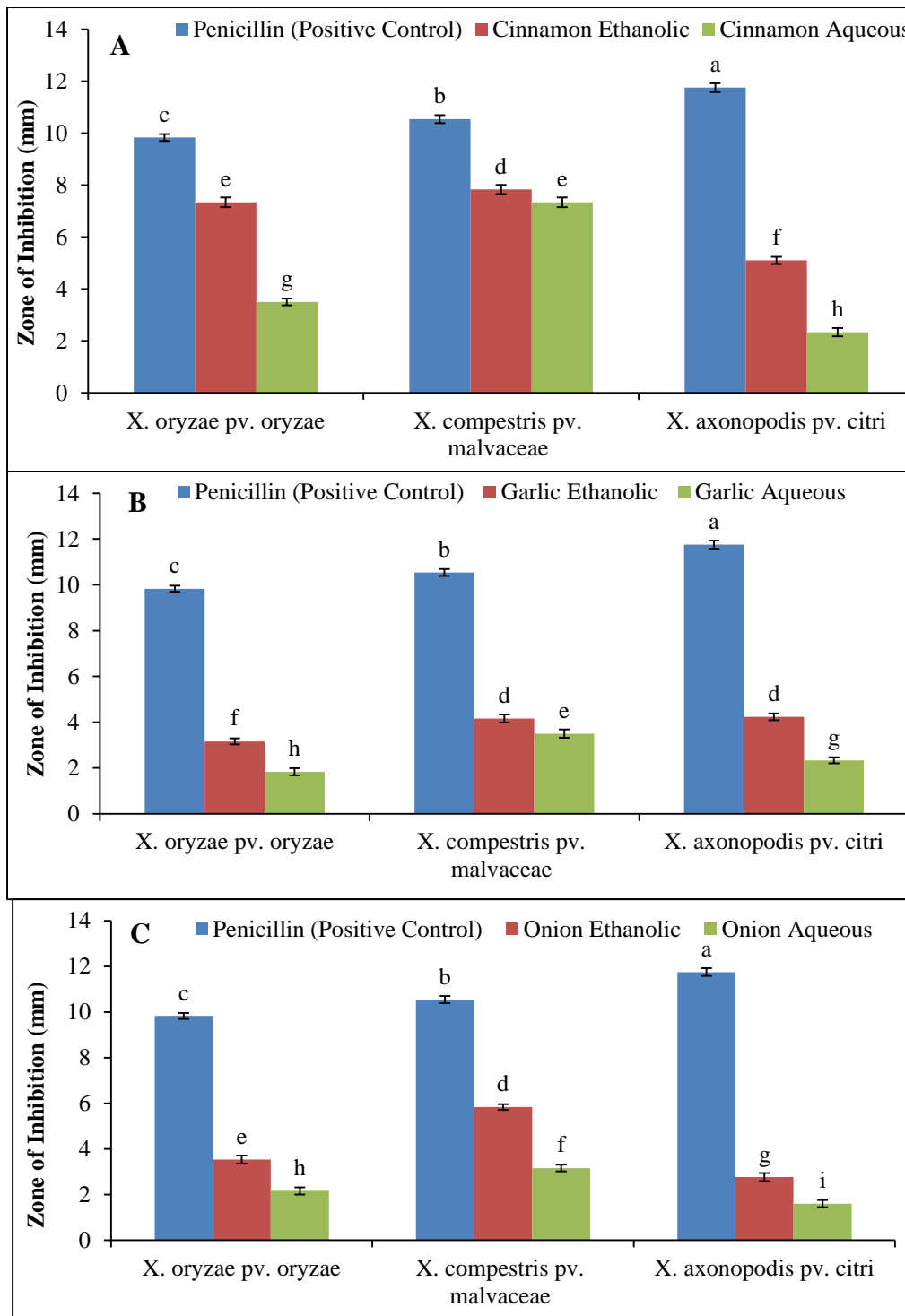


Fig. 1. Zone of inhibition of penicillin (positive control) and aqueous and ethanolic extracts of (A) Cinnamon, (B) Garlic and (C) Onion before growth of *Xanthomonas oryzae* pv. *oryzae*, *Xanthomonas campestris* pv. *malvacearum* and *Xanthomonas axonopodis* pv. *citri*. Values are mean \pm Standard Deviation. Letters a, b, c, d, etc. show significantly difference among different extracts

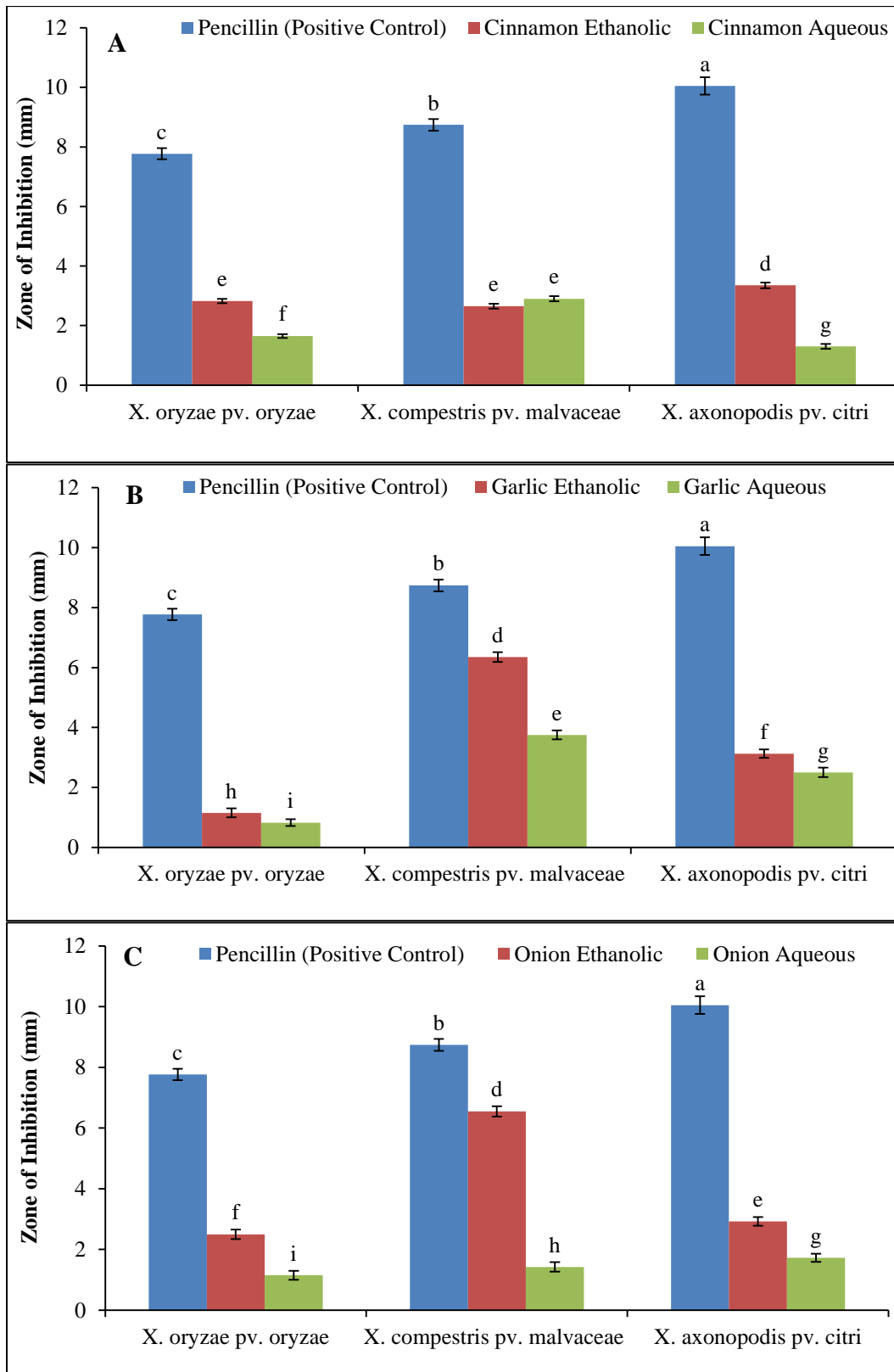


Fig. 2. Zone of inhibition of penicillin (positive control) and aqueous and ethanolic extracts of (A) Cinnamon, (B) Garlic and (C) Onion after growth of *Xanthomonas oryzae* pv. *oryzae*, *Xanthomonas campestris* pv. *malvacearum* and *Xanthomonas axonopodis* pv. *citri*. Values are mean \pm Standard Deviation. Letters a, b, c, d, etc. show significantly difference among different extracts

Table 1. MIC for bacterial pathogen

Sr. No.	Plant Name	Extract Type	<i>Xanthomonas campestris</i> pv. <i>malvacearum</i>			<i>Xanthomonas axonopodis</i> pv. <i>citri</i>			<i>Xanthomonas oryzae</i> pv. <i>oryzae</i>		
			Concentration mg/ml			Concentration mg/ml			Concentration mg/ml		
			66	33	0.01	66	33	0.01	66	33	0.01
1	Cinnamon	Ethanollic	3.5	1.5	0.0	2	1	0.0	5	2	0.0
		Aqueous	3	1	0.0	1.5	1	0.0	1.5	0.5	0.0
2	Onion	Ethanollic	3	1	0.0	2	0.5	0.0	2	0.5	0.0
		Aqueous	1.5	0.5	0.0	1	0.0	0.0	1	0.0	0.0
3	Garlic	Ethanollic	1.5	1	0.0	3	1	0.0	1.5	0.0	0.0
		Aqueous	1	0.0	0.0	2	0.5	0.0	1	0.0	0.0

3.3 Minimum Inhibitory Concentration (MIC)

As the concentration of extract decreases effectiveness of garlic, cinnamon, and onion also decreases to control growth of bacteria. All extracts used have minimum inhibitory concentration between 66 mg/ml to 33 mg/ml. Ethanolic and aqueous extracts of cinnamon showed maximum MIC against *Xanthomonas campestris* pv. *malvacearum* between 66 mg/ml to 33 mg/ml ranging from 3.5 mm to 1.5 mm and 3 mm to 1 mm, respectively. For *Xanthomonas axonopodis* pv. *citri* ethanolic (3 mm to 1 mm) and aqueous (2 mm to 0.5 mm) extracts of garlic showed maximum MIC (Table 1). However, for *Xanthomonas oryzae* pv. *oryzae* minimum inhibitory concentration was shown by ethanolic (5 mm to 2 mm) and aqueous (1.5 mm to 0.5 mm) extracts of cinnamon. Indu et al. [20] suggested that the effectiveness of garlic extract against different bacterial stereotypes can be due the presence of antibiotic compounds such as nalidixic acid.

The hydrogen and hydrophobic bonding of phenolic compounds with membrane proteins, disruption of the system of electron transport, membrane disintegration, and cell wall disintegration are all aspects of the antibacterial activity of different spices [21]. Anionic components including nitrate, thiocyanate, sulphates, and chlorides, as well as several other chemicals naturally found in plants, might be responsible for the antibacterial action of all these aqueous extracts [22]. When ethanolic extracts were assessed with aqueous extracts, the ethanol extract performed better because they dissolve larger organic molecules, leading to the discharge of more active antimicrobial substances [23].

4. CONCLUSION

In the present study, aqueous and ethanolic extracts of cinnamon, garlic and onion showed significant anti-bacterial activity against different species of *Xanthomonas* including *Xanthomonas campestris* pv. *malvacearum*, *Xanthomonas oryzae* pv. *oryzae*, *Xanthomonas axonopodis* pv. *citri*. Thus, it can be concluded that extracts of plants can control the growth of pathogenic bacteria and can be utilized to protect plants against pathogenic attacks in an environment-friendly manner.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Buttimer C, McAuliffe O, Ross RP, Hill C, O'Mahony J, Coffey A. Bacteriophages and bacterial plant diseases. *Frontiers in microbiology*. 2017;8:34.
2. Ryan RP, Vorhölter F-J, Potnis N, Jones JB, Van Sluys M-A, Bogdanove AJ, et al. Pathogenomics of *Xanthomonas*: understanding bacterium-plant interactions. *Nature Reviews Microbiology*. 2011;9(5): 344-55.
3. Poulin L, Grygiel P, Magne M, Gagnevin L, Rodriguez-R LM, Forero Serna N, et al. New multilocus variable-number tandem-repeat analysis tool for surveillance and local epidemiology of bacterial leaf blight and bacterial leaf streak of rice caused by *Xanthomonas oryzae*. *Applied and environmental microbiology*. 2015;81(2): 688-98.
4. Ebrahimi Z, Rezaei R, Masoumi-Asl A, Charehgani H. Variation in the aggressiveness of *Xanthomonas citri* subsp. *citri* pathotypes A and A* on three Citrus species, and epiphytic growth on some citrus weeds. *Crop and Pasture Science*. 2020;71(3):260-7.
5. Sandipan P, Patel R, Faldu G, Patel D, Solanki B. Relationship of bacterial leaf blight disease of cotton with different weather parameters under South Gujarat condition of India. 2018.
6. Arif M, Ali K, Jan MT, Shah Z, Jones DL, Quilliam RS. Integration of biochar with animal manure and nitrogen for improving maize yields and soil properties in calcareous semi-arid agroecosystems. *Field Crops Research*. 2016;195:28-35.
7. Islam W, Awais M, Noman A, Wu Z. Success of bio products against bacterial leaf blight disease of rice caused by *Xanthomonas oryzae* pv. *oryzae*. *PSM Microbiology*. 2016;1(2):50-5.
8. Elfarnini M, Abdel-hamid A, Achir M, Jamaledine J, Blaghen M. Antibacterial and antifungal activities of hexane and acetone extracts of sheets and fruits of Feijoa sellowiana O. *Biological Pharmaceutical Sciences*. 2018;3(1):35-44.

9. Martelli G, Giacomini D. Antibacterial and antioxidant activities for natural and synthetic dual-active compounds. *European Journal of Medicinal Chemistry*. 2018;158:91-105.
10. Gupta S, Didwania N. Role of medicinal plants as green pesticides against *Alternaria* blight. *Bulgarian Journal of Agricultural Science*. 2021;27(3):562-8.
11. Navidshad B, Darabighane B, Malecky M. Garlic: An alternative to antibiotics in poultry production, a review. *Iranian Journal of Applied Animal Science*. 2018;8(1):9-17.
12. Fratianni F, Riccardi R, Spigno P, Ombra MN, Cozzolino A, Tremonte P, et al. Biochemical characterization and antimicrobial and antifungal activity of two endemic varieties of garlic (*Allium sativum* L.) of the campania region, southern Italy. *Journal of medicinal food*. 2016;19(7):686-91.
13. Shrivastava A, Garg H. Allicin as a dermal antibiotic against microbial infections. *World Journal of Pharmaceutical Research* 2015;4(12):1052-6.
14. Sharma K, Mahato N, Lee YR. Systematic study on active compounds as antibacterial and antibiofilm agent in aging onions. *Journal of food and drug analysis*. 2018; 26(2):518-28.
15. Wang Y, Zhang Y, Shi Y-q, Pan X-h, Lu Y-h, Cao P. Antibacterial effects of cinnamon (*Cinnamomum zeylanicum*) bark essential oil on *Porphyromonas gingivalis*. *Microbial pathogenesis*. 2018;116:26-32.
16. Davis W, Stout T. Disc plate method of microbiological antibiotic assay: I. Factors influencing variability and error. *Applied microbiology*. 1971;22(4):659-65.
17. Wiegand I, Hilpert K, Hancock RE. Agar and broth dilution methods to determine the minimal inhibitory concentration (MIC) of antimicrobial substances. *Nature protocols*. 2008;3(2):163-75.
18. Bajpai M, Pande A, Tewari S, Prakash D. Phenolic contents and antioxidant activity of some food and medicinal plants. *International journal of food sciences nutrition*. 2005;56(4):287-91.
19. Jomehpour N, Ghazvini K, Jomehpour M. Antibacterial Activity of Aqueous and Methanolic Extracts of *Crocus sativus* Stigma and *Cinnamomum cassia* against Clinical Isolates of some Gram-Positive and Gram-Negative Pathogenic Bacteria. *Medical Laboratory Journal*. 2019;13(3): 31-4.
20. Indu M, Hatha A, Abirosh C, Harsha U, Vivekanandan G. Antimicrobial activity of some of the south-Indian spices against serotypes of *Escherichia coli*, *Salmonella*, *Listeria monocytogenes* and *Aeromonas hydrophila*. *Brazilian Journal of Microbiology*. 2006;37:153-8.
21. Souza ELd, Stamford TLM, Lima EdO, Trajano VN, Barbosa Filho JM. Antimicrobial effectiveness of spices: an approach for use in food conservation systems. *Brazilian Archives of Biology Technology*. 2005;48(4):549-58.
22. Darout IA, Christy AA, Skaug N, Egeberg PK. Identification and quantification of some potentially antimicrobial anionic components in miswak extract. *Indian Journal of Pharmacology*. 2000;32(1):11-4.
23. Cowan MM. Plant products as antimicrobial agents. *Clinical microbiology reviews*. 1999;12(4):564-82.

© 2021 Hussain et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:
<https://www.sdiarticle4.com/review-history/74590>