Journal of Pharmaceutical Research International



32(3): 9-14, 2020; Article no.JPRI.54968 ISSN: 2456-9119 (Past name: British Journal of Pharmaceutical Research, Past ISSN: 2231-2919, NLM ID: 101631759)

Chemo-typing of Three Egyptian Rosemary Oil for Their Chemical Composition and Anti-oxidant Activity

Menna I. Elshorbagy¹, Marwa Elsbaey², Hany N. Baraka^{1,2*} and Mohamed Farid Lahloub²

¹Department of Pharmacognosy, Faculty of Pharmacy, Delta University, Gamasa, Egypt. ²Department of Pharmacognosy, Faculty of Pharmacy, Mansoura University, Mansoura, Egypt.

Authors' contributions

This work was carried out in collaboration among all authors. Authors MFL and HNB designed the study, wrote the protocol and wrote the first draft of the manuscript. Authors ME and MIE managed the analyses of the study. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/JPRI/2020/v32i330409 <u>Editor(s):</u> (1) Dr. Q. Ping Dou, Karmanos Cancer Institute, School of Medicine, Wayne State University, USA. <u>Reviewers:</u> (1) Reginaldo dos Santos Pedroso, Universidade Federal de Uberlândia, Brazil. (2) Murad Ali Khan, Kohat University of Science and Technology, Pakistan. Complete Peer review History: <u>http://www.sdiarticle4.com/review-history/54968</u>

Original Research Article

Received 30 December 2019 Accepted 06 March 2020 Published 19 March 2020

ABSTRACT

Aims: To study the chemical composition of volatile oil samples from different place in Egypt (Mansoura, Gamsa and Assuit) and study their antioxidant activity determined by DPPH assay. **Study Design:** Hydro distillation of volatile oil samples and their GC/MS analysis and determination of their antioxidant activity by DPPH assay.

Place and Duration of Study: Department of Pharmacognosy, Faculty of pharmacy, Mansoura university, Egypt, between June 2015 and November 2017.

Methodology: The essential oil was isolated by hydro-distillation for 5 h using a Clevenger-type allglass apparatus according to the standard procedure of the European pharmacopeia and volatile oils analysis was performed by GC and GC-MS. GC analysis was carried out using Focus-DSQ-II GC/MS instrument (Thermo Scientific, MA, USA) equipped with TR-5 fused silica column (30 m × 0.25 mm, film thickness 0.25 μ m).

Results: The yield of the essential oils of three rosemary plants growing in Mansoura (RM), Gamsa (RG) and Assiut (RA) were 0.20% v/w, 0.32% v/w and 0.24% v/w, respectively. Their chemical

composition was analyzed by GC/MS, RM and RG were found to be α -pinene dominated chemotypes, 32.4% and 29.6%, respectively, meanwhile RA was camphor dominated chemotype (17.2%). Furthermore, their antioxidant activity was determined by DPPH assay. Their IC₅₀ values of essential oils of RM, RG and RA were 8.66 ± 0.7, 8.18 ± 0.5 and 9.74 ± 0.2, respectively. **Conclusion:** The GC/MS spectral data revealed the considerable difference between the chemical composition of essential oil constituents of RM, RG and RA which lead to different chemotypes. The present results also demonstrate that REO obtained from different areas in Egypt exhibited free radical scavenging activity determined by DPPH assay due to the synergistic effect between their constituents.

Keywords: Rosmarinus officinalis L.; essential oil; GC/MS; anti-oxidant activity.

ABBREVIATIONS

- GC/MS : Gas Chromatography and Mass Spectroscopy
- DPPH : 2.2 -Diphenyl-1-picryl-hydrazyl
- IC : Inhibitory Concentration
- RI : Retention Index
- BHT : Butyl Hydroxy Toluene
- REO : Rosemary Essential Oil
- RM : Essential Oil Sample of Mansoura
- RG : Essential Oil Sample of Gamsa
- RA : Essential Oil Sample of Assuit

1. INTRODUCTION

Rosmarinus officinalis L. is a very important aromatic and medicinal plant, which belongs to the Lamiaceae family [1]. It grows primarily in the Western Mediterranean region and it is now cultivated all over the world as aromatic and ornamental plant [2].

Essential oils are a complex mixture of volatile compounds with strong odor. They are synthesized in several plant organs and possess various biological activities [3].

Rosemary essential oils (REO) are commonly used as a condiment, but it has been widely used in the treatment of gastritis, dermatitis, inflammation and bronchitis, also used as antioxidant, anticancer and antimicrobial agents [4].

Chemotype describes the species of plant have the same morphological characters, but produce different quantities of the chemical components in their essential oil [5]. REO chemical composition differed significantly but the major constituents are 1,8 cineole, α -pinene, camphor, limonene, camphene and linalool [6], associated with variable amounts of other compounds such as borneol, terpinol and verbenone [2]. REO composition is affected by various factors, such as extraction method, the distance between plants, harvest time, soil humidity and drying method. Therefore, researchers have sought to obtain information about the yield, composition, and chemical properties of REO following a variety of extraction methods and harvesting times [7].

More than 13 different rosemary oil chemotypes have been reported based on the relative percentages of α -pinene, 1,8-cineole, camphor, borneol, verbenone and bornyl acetate [8].

The aim of the study is to determine the different chemotypes of the essential oil of various rosemary plants growing in Egypt. Rosemary plants were collected from Mansoura, Gamsa and Assiut. Mansoura and Gamsa lie on the east bank of the Damietta branch of the Nile, about 56 kilometers from each other, in the delta region. They are about 120 kilometers northeast of Cairo. Meanwhile, Assiut is located in Upper Egypt and lies about 375 kilometers south of Cairo. The essential oils obtained were investigated for their yield, chemical composition and antioxidant activity.

2. MATERIALS AND METHODS

2.1 Plant Material

Rosmarinus officinalis L. plants were collected from the botanical garden of Mansoura university, Mansoura (RM), botanical garden of delta university, Gamsa (RG) and botanical garden of Assuit University, Assuit (RA) in June 2015. A voucher specimen has been deposited at the herbarium of the Pharmacognosy Department, Faculty of Pharmacy, Mansoura University (06-15-RM-Mansoura), (06-15-RG-Mansoura) and (06-15-RA-Mansoura). Samples were kindly identified by Prof. Ibrahim Mashaly, Department of Ecology, Faculty of Science, Mansoura University, Egypt.

2.2 Distillation of the Essential Oil

Two hundred and fifty grams of the fresh aerial parts of each *R. officinalis* sample was chopped into small pieces. The essential oil was isolated by hydro-distillation for 5 h using a Clevenger-type all glass apparatus according to the standard procedure of the European pharmacopoeia [9]. Each oil was transferred to a screw-capped glass vial, dried over anhydrous sodium sulfate and stored at 4° in the dark until GC/MS analysis and biological activity screening.

2.3 GC/MS Analysis of Essential Oil Content

Analysis of volatile oils by GC-MS. GC analysis was carried out using Focus-DSQ-II GC/MS instrument (Thermo Scientific, MA, USA) equipped with TR-5 fused silica column (30 m × 0.25 mm, film thickness 0.25 µm). This work has been done in Spectroscopy Lab, Faculty of Science, Mansoura university. Sample volume; 10 µl. Oven temperature was programmed from 60°C to 240°C at 5°C /min; injector temperature, 275°C; carrier gas, helium (1 ml/min); split: 1/50. The relative proportions of the essential oil constituents were expressed as percentages obtained by peak area normalization. The MS operating parameters were: interface temperature: 300°C, ion source temperature: 200°C, El mode: 70 eV, scan range: 40 - 450 m/z. Compounds Identification: Mass spectra of the individual GC peaks were identified by computer search of the commercial libraries (WILEY, NIST), as well as matching with published mass spectra [10]. The identification was further confirmed by the calculation of the retention indices (RI) relative to n-alkanes [10]. The results were listed in Table 1.

2.4 DPPH (2.2 -Diphenyl-1-Picryl-Hydrazyl) Assay

The antioxidant activity was evaluated by the DPPH (1,1- diphenyl-2-picrylhydrazyl) radical method. The ability of the essential oils and main compounds to donate an electron and scavenge DPPH radical was determined by the method of Ojeda-Sana et al. [11]. Briefly, 10 μ L of each sample in triplicate and 120 μ L of DPPH solution in 70% methanol to start concentration of each sample of RM, RG and RA with 11.6 mg/ml, 18.3 mg/ml and 17.6 mg/ml, respectively. They were

added to the well flat-bottom microtitration plate. A DPPH° solution was used as blank sample and Butyl hydroxy toluene (BHT, Sigma Aldrich, Egypt), synthetic antioxidant was used as control. The plate was covered and incubated on a shaker for 1 h at 50 rpm at room temperature. Samples were read in a plate reader (DTX 880 Multimode detector, Beckman Coulter) using a 517 nm filter. The DPPH solution employed was freshly prepared, and stored protected from light. The antioxidant activity of the tested samples, expressed as percentage inhibition of DPPH, was calculated according to the formula IC (%) = [(A0 - A)/A0] x 100, where A0 and A are the absorbance values of the blank sample and the test, respectively. Percent inhibition after 30 min was plotted against concentration, and a linear regression was applied to obtain the IC₅₀ value [11] The results were recorded in Table 2.

3. RESULTS AND DISCUSSION

3.1 Essential Oils Analysis

The yield of essential oils of RM, RG and RA obtained by steam distillation were 0.20% v/w, 0.32% v/w and 0.24% v/w, respectively. Their chemical composition was determined by GC/MS analysis and the results were listed in (Table 1). Eleven compounds were identified from RM. representing 91.25 %, fourteen compounds from RG, representing 96.26% and seventeen compounds from RA, representing 95.59 % of the total detected constituents. The major constituents in the essential oils were α -pinene (32.45%, 29.65%) followed by1.8-cineole (22.12%. 12.57%) and camphor (11.67%, 11.86%) in RM and RG, respectively. While in RA, camphor (17.24%) was the major constituent followed by α -pinene (14.84%) and 1,8-cineole (11.50%).

The results also showed that the major constituents in the three essential oils samples are monoterpenes while sesquiterpenes were minor, representing 6.27%, 4.01% and 0.75% in RM, RG and RA, respectively.

The dominance of 1.8-cineole over any other constituent in rosemary plant is by the data reported by several authors. For example; Jean Claude Chalchat *et al.*, reported that 1,8-cineole was the major component in the essential oils of *Rosmarinus officinalis* L. collected from Morocco and Tunisia [8]. Where in Spain, Mehmet Musa Ozcan et al., reported that α -pinene was the major component in the essential oils of

Rosmarinus officinalis [5]. Myrcene was also found as the main component in aerial parts of *Rosmarinus officinalis* L. cultivated in South America [11]. For instance, a study conducted by Robert Tisserand and Rodney Young, 2014 revealed that the prominent component of the essential oil of *Rosmarinus officinalis* L. cultivated in France was bornyl acetate [12]. For interest, verbenone was reported as the major component for *Rosmarinus officinalis* L. oils cultivated in Egypt [12]. However, this is the first report of α -pinene and camphor dominated chemotypes form Egypt.

Our study showed that limonene (7.44%) and dihydrocarveol (1.18%) were present only in RA, so it was worth to mention that the presence of limonene and dihydrocarveol constituents is good evidence for essential oils of RA chemotype. On another hand, terpinene constituent (0.61%) was characteristic for the essential oils of RG only. α -Pinene (32.45 %) and 1.8-cineole (22.12%) were present with the highest percentage in RM over than RG and RA. While borneol (13.81 %) was the highest in RA more than RM and RG.

3.2 Antioxidant Activity

In the DPPH assay, the ability of the components of the essential oil to act as donors of hvdroaen atoms or electrons for the transformation of DPPH° radical into its reduced form DPPH was investigated. The essential oils studied were able to change the stable violet DPPH° radical into yellow-coloured DPPH, reaching 50% of reduction with IC_{50} values to become 8.66 \pm 0.7 µL/mL, 8.18 \pm 0.5 μ L/mL and 9.74 ± 0.2 μ l/ml for RM, RG and RA, respectively (Table 2). Our results are in agreement with previous data reported on the antioxidant activity of essential oils of rosemary [13].

No	Compound	RI	Percent co	Percent composition (%) of oil		
			Mansoura (RM)	Gamsa (RG)	Assiut (RA)	
1	α-Pinene	921	32.45	29.65	14.84	
2	Camphene	935	5.04	4.80	6.49	
3	α-Myrcene	968	-	-	1.20	
4	3-Carene	985	1.75	2.84	1.68	
5	Limonene	1010	-	-	7.44	
6	Eucalyptol	1025	22.12	12.57	11.50	
	(1,8-cineole)					
7	Terpinene	1052	-	0.61	-	
8	Terpinolene	1076	-	1.83	0.50	
9	α-Linalool	1094	-	2.91	5.46	
10	Camphor	1140	11.67	11.86	17.24	
11	3-Pinanone	1152	1.30	-	0.82	
12	Borneol	1164	2.41	8.19	13.81	
13	Terpinol	1174	-	1.05	3.18	
14	Verbenone	1201	4.89	5.74	7.93	
15	Dihydrocarveol	1265	-	-	1.18	
16	Isobornyl acetate	1287	3.35	10.20	1.57	
17	β-Caryophyllene	1416	5.59	3.40	0.34	
18	Caryophyllene oxide	1582	0.68	0.61	0.41	
Total identified constituents			91.25%	96.26%	95.59%	
-Hydr	rocarbon:					
	.83 % (RM), 43.13% (RG), 32	.49% (RA)				

Table 1. Composition of essential oil samples from R.	officinalis L.
---	----------------

-Monoterpene = 72.88% (RM), 65.1% (RG), 65.51%

(RA)

-Sesquiterpene = 6.27% (RM), 4.01 % (RG), 0.75% (RA)

-Total Oxygenated compounds =

46.42% (RM), 53.13% (RG), 63.1% (RA)

Bold values are the major constituents in the essential oils, RI= Retention index, (-) absent

Sample name	IC ₅₀ (μg/ml)		
R. officinalis of Mansoura (RM)	8.66 ± 0.7		
R. officinalis of Gamsa (RG)	8 .18± 0.5		
R. officinalis of Assuit (RA)	9.74 ± 0.2		
BHT	1.33 ± 0.08		

Table 2. Antioxidant IC₅₀ values of essential oils from of *R. officinalis* L.

Antioxidants minimize oxidation of the lipid components in foods and there is an increasing interest in the use of natural antioxidants in food preservation. BHT was used as a reference since it possesses well-known antioxidant properties [13]. In mammalian cells, REO only has not proved to be mutagenic, but also it was shown to have antimutagenic properties [14]. Therefore, our results suggest that REO can be a suitable target in the search for (natural) replacements for (synthetic) antioxidant food additives.

Our study revealed that α -pinene (32.45%, and 29.65%) was the essential oil major component in RM and RG, respectively. On the other hand, Camphor (17.24%) was the major component in RA. Although the essential oils of RM, RG and RA were different chemotypes, they had comparable antioxidant activities. We suggest that the antioxidant activity may be due to a synergistic effect between their constituents.

4. CONCLUSION

In the foregoing study, the GC/MS spectral data revealed considerable difference between the chemical composition of essential oil constituents of RM, RG and RA which lead to different chemotypes. They were revealed as α -pinene and camphor dominated chemotypes, which are reported for the first time from Egypt. The present results also demonstrate that REO obtained from different areas in Egypt exhibited free radical scavenging activity determined by DPPH assay due to the synergistic effect between their constituents. So, these results showed that rosemary may allow lowering the dose of synthetic antioxidants agents in foods.

CONSENT

It is not applicable.

ETHICAL APPROVAL

It is not applicable.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Stefanovits-bányai E, Tulok MH, Hegedûs A, Renner C, Varga IS. Antioxidant effect of various rosemary (*Rosmarinus* officinalis L.) clones. *Acta* Biologica Szegediensis. 2003;47:111–13.
- Rašković AL, Milanović I, Pavlović N, Ćebović T, Vukmirović S, Mikov M. Antioxidant activity of rosemary (*Rosmarinus officinalis* L.) essential oil and its hepatoprotective potential. BMC Complementary and Alternative Medicine. 2014;(14):225. Available:http://www.biomedcentral.com/1

472-6882/14/225

- Amin T, Naik HR, Hussain SZ. Chemotyping the essential oil in different rosemary (*Rosmarinus officinalis* L.) plants grown in Kashmir Valley. Biosciences, Biotechnology Research Asia. 2017; 14(3):1025–31
- Neito G, Ros G and Castillo J. Biological activities of three essential oils of the Lamiaceae Family. Medicines. 2017;4(4):63. Available:https://doi.org/10.3390/medicines 4030063
- Ozcan MM, Chalchat JC. Chemical composition and anti fungal activity of rosemary (*Rosmarinus officinalis* L.) oil from Turkey. International Jornal of Food Sceience and Nutrition. 2008;691-98. Available:https://doi.org/10.1080/09637480 701777944
- Hussain AI, Anwar F, Ali S, Chatha S, Jabbar A, Mahboob S, Nigam PS. *Rosmarinus officinalis* essential oil: Antiproliferative, antioxidant and antibacterial activities. Brazilian Journal of Microbiology. 2014;41(4):1070–78. Available:https://doi.org/10.1590/S1517-83822010000400027

Elshorbagy et al.; JPRI, 32(3): 9-14, 2020; Article no.JPRI.54968

- Bousbia N, Vian MA, Ferhat MA, Petitcolas E, Meklati BY, Chemat F. Comparison of two isolation methods for essential oil from rosemary leaves: Hydrodistillation and microwave hydrodiffusion and gravity. Food Chemistry. 2009;114(1):355–62. Available:https://doi.org/10.1016/j.foodche m.2008.09.106
- Chalchat JC, Garry RP, Michet A, Benjilali B, Chabart JL. Essential oils of rosemary (*Rosmarinus officinalis* L.). the chemical composition of oils of various origins (Morocco, Spain, France). Journal of Essential Oil Research. 1993;(6):613–18. Available:https://doi.org/10.1080/10412905 .1993.9698293
- Strasbourg, 7th ed. European directorate for the quality of medicines & healthcare (EDQM). European Pharmacopoeia; 2013.
- Adam RP. 4th ed, Allured Books, IL, USA. Identification of essential oil components by gas chromatography/mass Spectrometry; 2009.

- Ojeda- Sana AM, van Baren CM, Elechosa MA, Juárez MA, Moreno S. New insights into antibacterial and antioxidant activities of rosemary essential oils and their main components. Food Control 31. 2013;(1):189–95. Available:https://doi.org/10.1016/j.foodcont .2012.09.022
- 12. Tisserand R, Young R. 2nd ed. essential oil safety. A guide for health care professionals; 2014.
- Sebranek JG, Sewalt VJH, Robbons KLF, Houser TA. Comparison of natural rosemary extract and BHA/BHT for relative antioxidant effectiveness in pork sausage. Meat Science. 2005;289-296.
- Wang W, Li N, Zu Y, Efferth T. Antibacterial activity and anticancer activity of *Rosmarinus officinalis* L. essential oil compared to that of its main components. Molecules. 2012;17:2704-2713.

© 2020 Elshorbagy et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: http://www.sdiarticle4.com/review-history/54968