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In vitro Evaluation of Fungicides against Anthracnose of Betelvine (*Piper betle* L.)

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Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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ABSTRACT

Background: Betelvine is important commercial crop and the most profitable among all cultivated crops, which plays a vital role in the overall livelihood security of farm families. Diseases are the major yield constraints of crop plants. One of the most serious fungal diseases of dragon fruit is anthracnose caused by Colletotrichum species. Since less information available on anthracnose of betel vine, this study was undertaken.
Methods: The efficacy of non-systemic, systemic and combination fungicides were tested against Colletotrichum gloeosporioides using poisoned food technique (Vincent 1947) under in vitro condition. Six non-systemic fungicides Chlorothalonil 75% WP, Captan 50% WP, Mancozeb 75%

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WP, Copper oxychloride 50% WP, Propineb 70% WP and Copper hydroxide 53.8% at (250 ppm, 500 ppm and 1000 ppm), six systemic fungicides Hexaconazole 5% EC, Propiconazole 25 % EC, Azoxystrobin 25% SC, Tebuconazole 25.9% EC, Difenaconazole 25% EC and Picoxystrobin 22.5% SC at (100ppm, 150ppm, 250ppm) and six combi fungicides Propiconazole 13.9% + Difenoconazole 13.9% EC, Tebuconazole 50% + Trifloxystrobin 25% WG, Fluropyram 200 g/L + Tebuconazole 200 g/L SC, Fluxopyroxad 250 g/l + pyraclostrobin 250g/L, Fluopyram 250 g/L + Trifloxystrobin 250 g/L SC, Azoxystrobin 16.7% + Tricyclazole 33.3% SC at (150 ppm, 250 ppm, 500 ppm) were evaluated.

Results: Among six non-systemic fungicides evaluated against *C. gloeosporioides* which was obtained from the isolated sample and results revealed that the Copper hydroxide gave 69.90 % inhibition which was superior over all other fungicides evaluated and least inhibition was recorded with Mancozeb 40.30%. Difenconazole, Tebuconazole were the best systemic fungicides found best with inhibition % of 98.28 and 95.17 when evaluated against *C. gloeosporioides*. Out of the six evaluated combination products, propiconazole + difenconazole exhibited the highest inhibition rate at 99.78 %. Following closely, Fluopyram 200 g/L + Tebuconazole 200 g/L SC and Tebuconazole 50% EC + Trifloxystrobin 25% WG displayed inhibition rates of 89.47% and 87.50 % respectively.

Keywords: Anthracnose; betelvine; Colletotrichum gloeosporioides; fungicide.

1. INTRODUCTION

Betelvine (*Piper betle* L.), widely known as “paan" in the Indian sub-continent, has a long ancient history in India and occupies a significant place in the everyday life of the people as it is used in rituals and as medicine to cure many diseases and disorders. Malaysia is the most probable place of origin of the Betelvine [1]. “It belongs to the family Piperaceae. Betelvine is a perennial, dioecious, shade-loving, aromatic, evergreen root climber with glossy heart-shaped leaves and white catkin” [2]. “It is mainly grown in the tropics and subtropical regions, for its leaves are used as a chewing stimulant. In India, betelvine is grown throughout the country and as an important cash crop in southern parts, mainly in Andhra Pradesh, Karnataka, Kerala, and Tamil Nadu. Betel vine is also cultivated in Assam, Bihar, Madhya Pradesh, Maharashtra, Orissa, Tripura, Uttar Pradesh, and West Bengal with an estimated area of 53,539 ha” [3]. It is the most important cash crop, and that adequately justifies its nomenclature as the "Green gold of India" [4]. “Betelvine leaves and areca nut are used in many occasions like Hindu religious ceremonies, wedding ceremonies, and pujas. Chewing of pan leaf is an ancient habit that has existed for more than 2000 years” [5]. The essential diseases challenging betel leaf production are foot rot complex (*Phytophthora* spp, *Sclerotium rolfsii*, *Rhizoctonia solani*, *Macrophomina phaseolina*, and *Pythium vexans*), anthracnose (*Colletotrichum* spp.), leaf rot (*Colletotrichum* spp.), Powdery mildew (*Oidium piperis*) and bacterial leaf spot (*Xanthomonas betlicola*). The symptoms of betel vine anthracnose disease on the stem caused by *Colletotrichum* spp. at first appear as tiny, black, circular specks on the green bark of the stem. If conditions are dry, then these specks usually do not increase in size and remain as a black stain on the surface of the stem. The leaf spot disease appears only after the rain and affects only betelvine leaves. The disease infection will not spread to the vine. Environmental factors such as temperature, rainfall, relative humidity, and shade in baroja play vital roles in the disease development. The high relative humidity (92 %) was critical for severe leaf spot disease and led to heavy loss in betel vine crops [6].

2. MATERIALS AND METHODS

The poisoned food technique [7], was followed to evaluate the efficacy of non-systemic, systemic fungicides and combi products in inhibiting the mycelial growth of pathogen. The fungus was grown on potato dextrose agar medium for 12 days prior to setting up the experiment. The potato dextrose agar medium was prepared and melted with the use of microwave oven. The fungicidal suspension was added to the melted medium to obtain the required concentrations on commercial formulation basis of the fungicide. 20 ml of poisoned media was poured in each sterilized Petri plates. Control treatment was maintained without addition of fungicide. Mycelial disc of 5 mm was taken from the periphery of 12 days old colony was placed in the center of Petri plates and incubated at 27 ± 1ºC for 12 days and three replications were maintained for each treatment. The diameter of the colony was measured in two directions and average was

recorded. Percent inhibition mycelial growth of the fungus was calculated by using the formula given by Vincent [8].

$$I = \frac{C - T}{C} \times 100$$

Where,

I = Percent inhibition
C = Radial growth in control
T = Radial growth in treatment (fungicide)

3. RESULTS AND DISCUSSION

The assessment of fungicides through *in vitro* testing proves to be a convenient method for assessing a substantial array of chemicals, gauging their effectiveness in restraining pathogen growth. This approach swiftly furnishes valuable initial insights into the fungicides' potency against the pathogen, offering a concise timeframe for evaluation. These findings then act as a compass for subsequent field trials. In this ongoing study, a total of six contact, six systemic, and six combined fungicide products were examined against *Colletotrichum gloeosporioides*, encompassing three distinct concentrations.

Among six contact fungicides evaluated against *C. gloeosporioides* and Copper hydroxide gave 69.90% inhibition which was superior over all other fungicides evaluated. Which was followed by Copper oxychloride (62.96%), Chlorothalonil (53.87%), Propineb (47.85%), Captan (40.83%) and least inhibition was recorded with Mancozeb 40.30% (Fig 1).

The results were similar with work of Parvathy and Girija, [9]. Copper based fungicides are effective because it kills the pathogen by denaturing proteins and enzymes in cells of pathogens when they come in contact.

Among six different systemic fungicides evaluated against *C. gloeosporioides* Difenconazole, Tebuconazole were found best with inhibition percentage of 98.28 and 95.17. These results were similar to the earlier reports made by Prashanth et al. [10], Ahmed et al. [11], Parvathy and Girija, [9] that Difenconazole and Tebuconazole have highest inhibition percentage on the growth of *C. gloeosporioides*. (Fig 2).

The effectiveness of the Triazole fungicides may be attributed to their interference with the biosynthesis of fungal sterols and inhibit the ergosterol biosynthesis. In many fungi, ergosterol is essential for the structure of cell wall and its absence cause irreparable damage to cell wall leading to death of fungal cell. A similar study was reported for the effectiveness of Triazoles, which inhibit the sterol biosynthesis pathway in fungi [7].

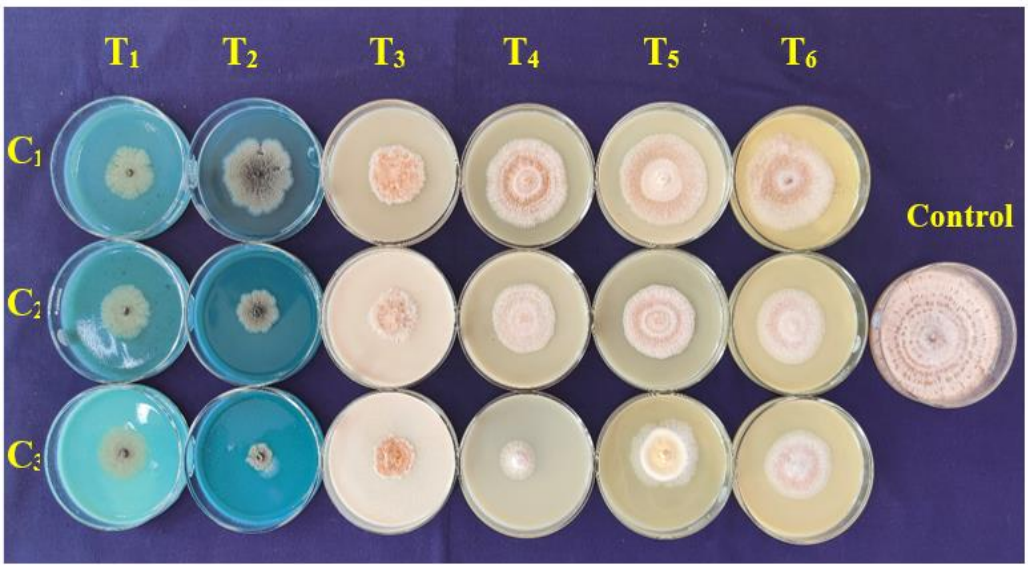

Fig. 1. *In-vitro* evaluation of non-systemic fungicides against Colletotrichum gloeosporioides
T1: Copper oxychloride; T2: Copper hydroxide; T3: Propineb; T4: Chlorothalonil; T5: Captan; T6: Mancozeb; C1: 250 ppm; C2: 500 ppm; C3: 1000 ppm

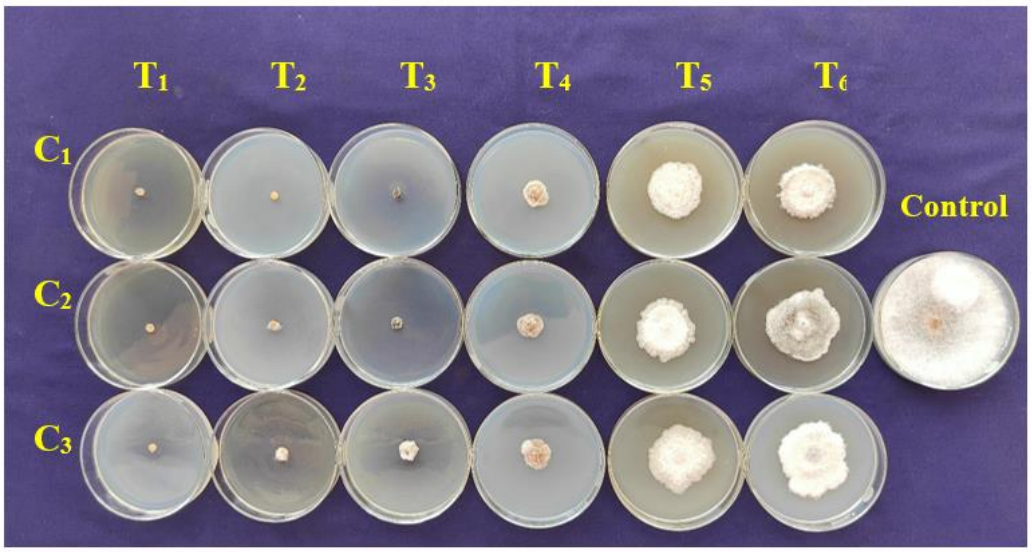

Fig. 2. *In-vitro* evaluation of systemic fungicides against *Colletotrichum gloeosporioides*

T1: Difenoconazole; T2: Tebuconazole; T3: Propiconazole; T4: Hexaconazole; T5: Picoxystrobin; T6: Azoxystrobin; C1: 250 ppm; C2: 150 ppm; C3: 100 ppm

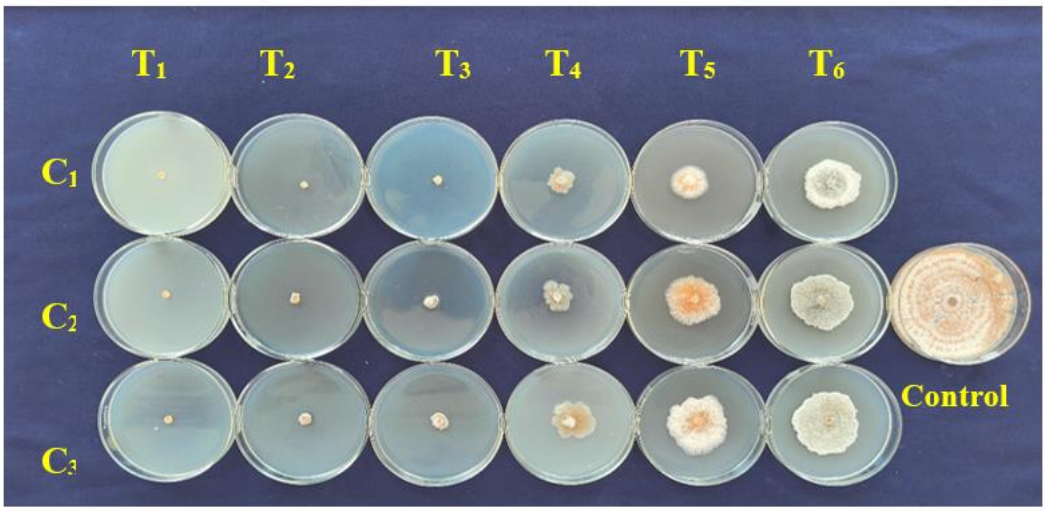

Fig. 3. *In-vitro* evaluation of combi products against *Colletotrichum gloeosporioides*

T1: Propiconazole + Difenoconazole; T2: Fluopyram + Tebuconazole; T3: Tebuconazole + Trifloxystrobin; T4: Fluxapyroxad + Pyraclostrobin; T5: Azoxystrobin + Tricyclazole; T6: Fluopyram + Trifloxystrobin; C1: 500 ppm; C2: 250 ppm; C3: 150 ppm

Out of the six evaluated combination products, propiconazole + difenconazole exhibited the highest inhibition rate at 99.78%. Following closely, Fluopyram 200 g/L + Tebuconazole 200 g/L SC and Tebuconazole 50% EC + Trifloxystrobin 25% WG displayed inhibition rates of 89.47% and 87.50% respectively. These findings aligned with the research by Prashanth et al. [10], Parvathy and Girija, [9], and Pavithra and Benagi, [12]. (Fig 3).

The utilization of combination fungicides effectively curbs the development of fungal resistance to systemic fungicides. This is because systemic fungicides disrupt only a single, or occasionally two, functions within the fungal physiology, which can be easily overcome by a singular mutation. On the contrary, non-systemic protectant fungicides impact numerous aspects of fungal physiology, requiring the fungus to undergo multiple changes in order to

develop resistance. As a result, the combination of both systemic and non-systemic fungicides yields superior outcomes.

4. CONCLUSION

In-vitro efficacy of non-systemic, systemic and combi fungicides done to know their efficiency in suppressing the growth of *C. gloeosporioides* revealed the efficacy of copper hydroxide which showed 69.90 per cent inhibition. In case of systemic fungicides maximum per cent inhibition was recorded in Difenoconazole (98.28%) and lowest per cent inhibition by Azoxystrobin (48.18%). While among combi products Propiconazole 13.9% EC + Difenoconazole 13.9% EC showed 99.78% followed by Fluopyram 200 g/l + Tebuconazole 200 g/l SC (89.47%), Tebuconazole 50% EC + Trifloxystrobin 25% WG (87.50%).

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Chattapdayay SP, Maity S. Diseases of betelvine and species. ICAR, New Delhi; 1967.
2. Hiralal Jana, Debabrata Basu, Suchhanda Jana. Pesticides use pattern of betelvine growers in controlling diseases in Midnapur (east) district of West Bengal. International Journal of Current Advanced Research. 2016;6:5314-5320.
3. Ray DP. Keynote Address, National Seminar on Piperaceae, 21-22 November 2008, IISR, Calicut. 2008;26.
4. Nutankumar SJ, Deshmukh P. Madhavi SJ. Review of study of different diseases on betelvine plant and control measure. JAIEM. 2014;3(3):560 563.
5. Kumar V, Suryanarayana MAT, Bindu HK. Status of Betelvine Cultivation in Karnataka, Section of Medicinal Crops, Indian Institute of Horticultural Research, Bangalore. 2014;1- 4.
6. Dasgupta B, Sen C. Assessment of Phytophthora root rot of betelvine and its management using chemicals. Journal of Mycology and Plant Pathology. 1999;29:91-95.
7. Nene YL, Thapliyal PN. Evaluation of fungicides. In: Fungicides in plant disease control (3rd ed.) Oxford, IBM Publishing Co, New Delhi; 1993.
8. Vincent JM. Distribution of fungal hyphae in the presence of certain inhibitors. Nature. 1947;159: 850.
9. Parvathy R, Girija VK. In vitro evaluation of fungicides and organic preparations against Colletotrichum gloeosporioides causing anthracnose of black pepper (*Piper nigrum* L.). International Journal of Applied and Pure Science and Agriculture. 2016;2:2394-2398.
10. Prashanth A, Arun RS, Naik MK, Patil MB, Rajesh SP. Evaluation of fungicides, bio agents and botanicals against pomegranate anthracnose. Indian Journal of Plant Protection. 2008;36(2):283-287.
11. Ahmed MJ, Hossain KS, Bashar MA. Anthracnose of betel vine and it's *In vitro* management. Dhaka University Journal of Biological Sciences. 2014;23(2): 127-133.
12. Pavithra S. Benagi VI. In vitro evaluation of fungicides, botanicals and bio-agents against Colletotrichum gloeosporioides (Penz.) Penz. and Sacc. the causal agent of anthracnose of pomegranate. Environmental Ecology. 2017;35(2):671-675.

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