#### Advances in Bioscience and Biotechnology, 2015, 6, 501-507

Published Online August 2015 in SciRes. <a href="http://www.scirp.org/journal/abb">http://www.scirp.org/journal/abb</a> http://dx.doi.org/10.4236/abb.2015.68052



# Simultaneous Use of Entomopathogenic Fungus *Beauveria bassiana* and Diatomaceous Earth against the Larvae of Indian Meal Moth, *Plodia interpunctella*

## Mohsen Arooni-Hesari, Reza Talaei-Hassanloui\*, Qodrat Sabahi

Department of Plant Protection, College of Agriculture and Natural Resources, University of Tehran, Karaj, Iran Email: \*rtalaei@ut.ac.ir

Received 2 June 2015; accepted 31 July 2015; published 3 August 2015

Copyright © 2015 by authors and Scientific Research Publishing Inc.
This work is licensed under the Creative Commons Attribution International License (CC BY). <a href="http://creativecommons.org/licenses/by/4.0/">http://creativecommons.org/licenses/by/4.0/</a>



Open Access

# **Abstract**

The suppressive ability of entomopathogenic fungus *Beauveria bassiana* alone and in combination with diatomaceous earth (DE) was studied against the larvae of Indian meal moth, *Plodia interpunctella* (*Hübner*) (Lep., Pyralidae). This study clearly showed that simultaneous use of *B. bassiana* and DE against larvae of *P. interpunctella*, not only could reduce the required concentration of fungal conidia or DE, but also could shorten the time need for showing insecticidal effects. The  $LC_{50}$  value of fungus at 7 d after treatment was  $9.8 \times 10^5$  conidia  $mg^{-1}$  diet. Larvae showed a dose response to *B. bassiana*, and the addition of diatomaceous earth at 500 and 2000 ppm resulted in a significant increase in mortality. Larval mortality reached to the maximum of 28.3% and 71.7% after 7 d exposure to 500 and 2000 ppm DE concentrations, respectively. The  $LC_{50}$  value for *B. bassiana* in the presence of DE 500 ppm was  $4.6 \times 10^4$  con.  $mg^{-1}$  diet and of DE 2000 ppm was  $1.65 \times 10^3$  con.  $mg^{-1}$  diet. According to our results, *B. bassiana* and DE can be considered as two suitable candidates for integration into IPM strategy.

# **Keywords**

Stored Product Pest, Beauveria bassiana, Synergism, Diatomaceous Earth, Bioassay

## 1. Introduction

Entomopathogenic fungi are among the important biological control agents of insect pests, by causing lethal in-

\*Corresponding author.

How to cite this paper: Arooni-Hesari, M., Talaei-Hassanloui, R. and Sabahi, Q. (2015) Simultaneous Use of Entomopathogenic Fungus *Beauveria bassiana* and Diatomaceous Earth against the Larvae of Indian Meal Moth, *Plodia interpunctella*. *Advances in Bioscience and Biotechnology*, **6**, 501-507. <a href="http://dx.doi.org/10.4236/abb.2015.68052">http://dx.doi.org/10.4236/abb.2015.68052</a>

fections and regulating insect and mite populations in nature by epizootics [1]-[3]. These biocontrol agents infect a wide range of insect orders including Hemiptera, Coleoptera and Lepidoptera which are of great concern in worldwide agriculture but some strains of these fungi could be host specific with a very low risk of attacking non-target organisms or beneficial insects [4].

The entomopathogenic as comycete, *Beauveria bassiana* (Balsamo) Vuillemin is an important pathogen of insects and it has been developed as a microbial insecticide for use against many major arthropod pests [5] [6]. It has been developed as a microbial insecticide for use against many major pests, including lepidopterans. It is reported to be non-toxic to humans and other vertebrates, so it can be applied on commodities [7].

*Plodia interpunctella* (Hübner) (Lepidoptera: Pyralidae), is a serious pest of raw and processed food products worldwide [8]-[11]. Some reports have pointed out to the promising control effects of *B. bassiana* on Indian meal moth [12] [13]. However the fungi encountered some limiting factors to show adequate level of pest control.

The use of *B. bassiana* alone for the control of many stored product pests requires a high application rates. The action of fungal penetration to the host integument is dependent to physicochemical properties of cuticle including thickness, sclerotization, and presence of fatty acids as well as cuticle destroying enzymes [14]-[16].

The presence of cuticular lipids in insect integument could protect the insect from pathogenic microorganisms. If any agent could damage this barrier, greater penetration and higher pathogen virulence would be expected.

Diatomaceous Earth (DE), a natural product composed of the fossils of diatoms, can exhibit such role with its special properties like integument scarification and adsorbing wax from cuticle layer [17], causing the release of subcuticular compounds that have a synergistic effect on conidial attachment and virulence of entomopathogenic fungi [18] [19].

Several studies document that DE formulations are very effective against a wide range of stored products pests, persist on the product for a long period and can be easily removed from the grains [20]-[22].

It is generally accepted that these materials act as desiccants on insect cuticle, and that insects exposed to DE particles die from rapid water loss [17] [23] [24]. Low mammalian toxicity is another important characteristic of DE [25], rendering it a potential and safe method for pest control.

Our objective in this study was to determine the effects of *B. bassiana* and DE, alone or in combination, on the Indian meal moth larvae as well as clarify interactions of these two agents.

#### 2. Materials and Methods

#### 2.1. Insect Rearing

Indian meal moth, *P. interpunctella* was acquired from a laboratory population at the Department of Plant Protection, University of Tehran and reared under laboratory conditions ( $25^{\circ}C \pm 1^{\circ}C$ ,  $65\% \pm 10\%$  RH and 16:8 L:D) in Polyethylene boxes ( $30 \times 20 \times 15$  cm) covered with a piece of fine cloth.

Artificial diet including wheat bran, yeast, honey, and glycerol, prepared by the method described by Sait *et al.* [26], was used for insect rearing. No antibiotic or fungicide was added to the diet.

# 2.2. Fungal Isolate and Cultures

The isolate used in this study was *B. bassiana* ETU105, originally isolated from soil using *Galleria*-bait and preserved in Laboratory of Biological Control at the Department of Plant Protection, University of Tehran. Fungal isolate was grown on Sabouraud dextrose agar plus 1% yeast extract (SDAY) in 9 cm diameter Petri dishes (**Figure 1**) and incubated under conditions of 25°C  $\pm$  1°C, 75  $\pm$  10 RH and 16:8 L:D for 14 days. Colony was preserved at 4°C.

Fungal conidia were collected by scraping conidial layer using sterilized scalpel into 0.02% Tween 80. The conidial concentration was estimated using a haemocytometer. Each mg of collected conidia contained  $4.5 \times 10^8$  spores. Five serial concentrations  $10^3$  -  $10^7$  conidia mg<sup>-1</sup> diet were prepared by mixing adequate conidia with artificial diet. An electric mixer was used for 5 min to prepare an even mixture.

#### 2.3. Diatomaceous Earth

Diatomaceous Earth (DE) was prepared from Kimia Sabz Avar Company located in Iran. It was oven-dried initially at 60°C for two hours, and then two concentrations of 500 and 2000 mg/kg were prepared by adding sufficient DE to artificial diet. These two concentrations were selected based on the results of our preliminary test.

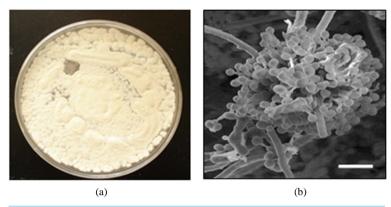


Figure 1. Beauveria bassiana, (a) Fully sporulated on SDAY medium; (b) Scanning electron micrograph of conidiophores and conidia clusters (original, bar = 10 μm).

## 2.4. Bioassay

There were 17 treatments: five fungal dose rates alone, five fungal dose rates in combination with each of the two DE dose rates and the two DE rates alone. Five serial dose rates of fungal conidia were  $10^3$  -  $10^7$  con.  $mg^{-1}$  diet. Fifteen g of diet containing treatments were placed in series of glass Petri dishes with 9 cm in diameter. An additional series of dishes containing untreated diet served as a control.

After the preparation of the dishes, 15 third instar larvae of *P. interpunctella* were introduced into each dish. The dishes were placed in incubators set up at temperature of  $25^{\circ}\text{C} \pm 1^{\circ}\text{C}$  and RH of  $65\% \pm 5\%$  for a week. All the experiments were repeated four times. Larval mortality was assessed daily until 7 days after exposure.

# 2.5. Statistical Analysis

Mortality counts were corrected by using Abbott's formula [27]. The experiment was designed and conducted in a factorial Complete Randomized Design (CRD). The pooled data were analyzed separately for each treatment category (Fungus alone, DE alone, DE 500+ fungus, DE 2000+ fungus), by submitting the mortality counts to ANOVA's for dose rate. Means were separated using the Tukey test at P < 0.05. SAS software version 9.1 was used for statistical analysis. The LC<sub>50</sub> values were estimated with probit analysis using POLO-Plus [28].

## 3. Results

# 3.1. Effect of B. bassiana on P. interpunctella Larvae

Mortality rates of third instar larvae of P. interpunctella treated with B. bassiana increased with increasing conidial concentration and time of exposure. Differences among lethal effects established by different conidial concentrations of fungal isolate were significantly different (F = 72.49, df = 5, P < 0.0001).

The LC<sub>50</sub> value of fungus at 7 d after treatment on *P. interpunctella* larvae was  $9.8 \times 10^5$  con. mg<sup>-1</sup> diet. Comparative virulence of *B. bassiana* isolate against L<sub>3</sub> instar of *P. interpunctella* indicated that mortality of larvae began at the third day after exposure and reached to 61.7% at the day 7. The LT<sub>50</sub> value of fungus at  $1 \times 10^7$  con. mg<sup>-1</sup> was 6.59 d (6.23 - 7.12 days lower and upper 95% C.I., respectively).

#### 3.2. Effect of Diatomaceous Earth on *P. interpunctella* Larvae

Larval mortality reached to a maximum of 28.3% and 71.7% after 7 d exposure to 500 and 2000 mg/kg DE concentrations, respectively. There were highly significant differences in larval mortality resulting from different concentrations of DE (F = 121.13, df = 2, P < 0.0001). Larval mortality ranged from 1.7 to a maximum of 71.7% over the range of tested concentrations.

# 3.3. Combination of B. bassiana and DE

Data on comparative virulence of B. bassiana isolate in combination with DE against L<sub>3</sub> instar of P. interpunctella

is brought in **Table 1**. These results showed that there was a synergistic relationship between fungus and DE targeting larvae of *P. interpunctella*.

The LC<sub>50</sub> value for *B. bassiana* plus DE 500 mg/kg was  $4.6 \times 10^4$  con. mg<sup>-1</sup> diet and with DE 2000 mg/kg was  $1.65 \times 10^3$  con. mg<sup>-1</sup> diet in contrast to the LC<sub>50</sub> of  $9.8 \times 10^5$  con. mg<sup>-1</sup> diet for *B. bssiana* alone (**Table 2**). There were significant differences among these treatments (F = 72.49, df = 5, P < 0.0001).

The combination treatment of fungus with DE caused the highest mortality at 7d after treatment which was significantly greater than all other treatments. With concentration of 500 ppm DE, mortality of larvae began at the second day after exposure to  $1 \times 10^7$  con. mg<sup>-1</sup> of fungus and reached to 100% at the day 7.

With DE 500, LT<sub>50</sub> value for fungus with concentration of  $1 \times 10^5$  con. mg<sup>-1</sup>, diminished to 3.9 d which was significantly lower than that of fungus alone with 6.6 d (F = 11.48, df = 26, P < 0.0001). This value showed a greater reduction when DE 2000 was applied, such that it was reduced to 2.1 d. The log-probit regression lines of LC<sub>50</sub> had slopes of 0.34 without DE, 0.65 with DE 500 and 0.64 with DE 2000. These parameters for LT<sub>50s</sub> were 8.21, 6.54 and 5.35, respectively (Table 3).

#### 4. Discussion

Results from bioassays on *P. interpunctella* larvae showed that the *B. bassiana* isolate ETU105 was virulent against the pest, however, a relatively high concentration of  $9.8 \times 10^5$  conidia mg<sup>-1</sup> diet needs to cause 50% mortality during 7 d. These results stand somewhat in agreement with that of Buda and Pečiulytė [29] who found that after treatment with concentration of  $2.6 \times 10^6$  conidia mg<sup>-1</sup> of *B. bassiana*, larval mortality of the species, was reached to 50% during 5 days.

Many of the important pests in grain storage have proven to be susceptible to *B. bassiana*, but its high production costs and high application rates make it economically unreachable [30]-[36].

**Table 1.** Mean percent mortalities of *P. interpunctella* third instar larvae caused by *B. bassiana* in combination with diatomaceous earth.

DE concentration (ppm)	Fungal concentration (conidia mg <sup>-1</sup> diet)							
	0	10 <sup>3</sup>	$10^{4}$	10 <sup>5</sup>	10 <sup>6</sup>	10 <sup>7</sup>		
0	5	25	28.33	43.33	55	61.66		
500	38.33	41.66	60	71.66	80	100		
2000	71.66	83.33	91.66	96.66	98.33	100		

Table 2. LC<sub>50</sub> values of *B. bassiana* alone or in combination with two diatomaceous earth doses based on experiments with  $3^{rd}$  instar larvae of *P. interpunctella*.

DE concentration (ppm)	Fungus LC <sub>50</sub> (con. mg <sup>-1</sup> diet)	$\chi^2$	Slope	Lower 95% confidence limit	Upper 95% confidence limit
0	$0.98 \times 10^{6}$	4.23	0.34	$0.28\times10^6$	$0.50~\times~10^7$
500	$4.06\times10^4$	8.86	0.65	$6.11 \times 10^{3}$	$0.12 \times 10^{6}$
2000	$1.65\times10^3$	8.21	0.64	$0.26\times10^2$	$1.03 \times 10^4$

Number of treated larvae = 360.

Table 3. Values of LT<sub>50</sub> for *B. bassiana* at  $1 \times 10^7$  con. mg<sup>-1</sup> diet alone or in combination with two doses of diatomaceous earth based on experiments with  $3^{rd}$  instar larvae of *P. interpunctella*.

DE concentration (ppm)	Fungus LT <sub>50</sub> (d)	$\chi^2$	Slope	Lower 95% confidence limit	Upper 95% confidence limit
DE 0	6.59	5.9	8.21	6.23	7.12
DE 500	3.92	9.68	6.54	3.69	4.15
DE 2000	2.09	8.08	5.35	1.90	2.28

Number of treated larvae = 360; df = 26.

The concentration of 500 ppm of DE, caused 28.3% mortality of *P. interpunctella* larvae with increasing the DE concentration to 2000 ppm could enhance the mortality rate to 71.7 percent. This result approved the finding of Sabbour *et al.* [13], however, the LC<sub>50</sub> of fungus in their study was  $1.29 \times 10^9$  con. mg<sup>-1</sup> which was much greater than that of our finding,  $1 \times 10^5$  con. mg<sup>-1</sup> diet. This difference may be related to fungus isolate or population of *P. interpunctella*.

Our results also are in conformity with findings of Akbar *et al.* [18] who found that DE increased the efficacy of *B. bassiana* against *Tribolium castaneum* larvae. Also Michalaki *et al.* [19] found that the effectiveness of entomopathogenic fungus, *Metarhizium anisopliae* can be benefitted by the presence of DE against *T. confusum* larvae.

DE does not leave any toxic residue because of its physical effects; however, regarding the possible health issues, lower concentrations should be preferred. Low concentration of DE does not leave any harmful residue on stored commodities. If it could be prepared from good quality and suitable resources, would have very low effect on human breathing system. Also, using suitable breathing mask, such effects could be greatly diminished.

Fast and stable effectiveness, make DE an appropriate choice to substitute chemical compounds for stored product protection. Moisture increase of environment can lead to less effectiveness of DE, so it is more successful in control of stored product pests. Moreover such pests are little in size, have a higher body surface to body volume ratio, and encountered to low water content of commodities. The lack of insect resistance against DE is another important prominence which enhances its capability for use as a part of integrated pest management program.

DE does not have any significant effect on stored products by own itself and can be removed easily with air blowing or washing. This can decrease applying chemical pesticides in grain deposit. Using DE at concentration of 500 ppm, the  $LC_{50}$  value of *B. bassiana* was significantly decreased on *P. interpunctella* larvae.

The exact mechanisms by which DE interacts with *B. bassiana* are not clear but may involve a combination of increased availability of water and other nutrients, removal or mitigation of inhibitory materials, alteration of adhesive properties, and physical disruption of the cuticular barrier [18]. Lord [12] [34] proposed that lipid removal may contribute to the synergistic interaction between *B. bassiana* and DE against some stored grain pests.

Akbar *et al.* [18] showed the number of *B. bassiana* conidia attached to the larval cuticle of *Tribolium castaneum* was significantly greater with DE presence than without it. The mean counts of conidia were 212.7 with DE and 90.9 without DE.

Michalaki *et al.* [19] revealed that DE benefits the fungal efficacy only when conidial concentration exceeds a certain "active threshold". Below this "threshold", a considerable amount of conidia may be damaged by the presence of DE, or DE particles may partially lose their desiccant capacity.

# 5. Conclusion

Finally, this study clearly indicated that simultaneous use of *B. bassiana* and DE against larvae of *P. interpunctella*, not only reduced the required concentration of fungus or DE, but it would also shorten the time need for showing their effects. On the other hand, the greatest factor in the loss of inoculum viability of entomopathogenic fungi under field conditions is inactivation caused by UV light [37] [38], but grain storage environment does not have such disadvantage, thus it could be a suitable environment for integrated application of *B. bassiana* and DE.

#### References

- [1] Burges, H.D. and Weiser, J. (1973) Occurrence of Pathogens of the Flour Beetle, *Tribolium castaneum. Journal of Invertebrate Pathology*, **22**, 464-466. <a href="http://dx.doi.org/10.1016/0022-2011(73)90178-X">http://dx.doi.org/10.1016/0022-2011(73)90178-X</a>
- [2] Carruthers, R.I. and Soper, R.S. (1987) Fungal Diseases. In: Fuxa, J. and Tanada, Y., Eds., Epizootiology of Insect Diseases, John Wiley and Sons Inc., New York, 357-416.
- [3] McCoy, C.W., Samson, R.A. and Boucias, D.G. (1988) Entomogenous Fungi. In: Ignoffo, C.M. and Mandava, N.B., Eds., *Handbook of Natural Pesticides*, *Vol. V, Microbial Insecticides*, *Part A*, CRC Press, Boca Raton, 151-236.
- [4] Roberts, D.W. and Humber, R.A. (1981) Entomogenous Fungi. In: Cole, G.T. and Kendrick, B., Eds., Biology of Conidial Fungi, Academic Press, New York, 201-236. http://dx.doi.org/10.1016/b978-0-12-179502-3.50014-5
- [5] Charnley, A.K. and Collins, S.A. (2007) Entomopathogenic Fungi and Their Role in Pest Control. In: Kubicek, C.P. and Druzhinina, I.S., Eds., *Environmental and Microbial Relationships*, Springer, Berlin, 159-187.
- [6] Faria, M.R. and Wraight, S.P. (2007) Mycoinsecticides and Mycoacaricides: A Comprehensive List with Worldwide Coverage and International Classification of Formulation Types. *Biological Control*, 43, 237-256.

- http://dx.doi.org/10.1016/j.biocontrol.2007.08.001
- [7] Mahroof, R. and Subramanyam, B. (2006) Susceptibility of *Plodia interpunctella* (Lepidoptera: Pyralidae) Developmental Stages to High Temperatures Used during Structural Heat Treatments. *Bulletin of Entomological Research*, 96, 539-545. http://dx.doi.org/10.1017/BER2006454
- [8] Sedlacek, J.D., Weston, P.A. and Barney, R.J. (1996) Lepidoptera and Psocoptera. In: Subramanyam, B. and Hagstrum, D.W., Eds., *Integrated Management of Insects in Stored Products*, Marcel Dekker Inc., New York, 63-66.
- [9] Doud, C.W. and Phillips, T.W. (2000) Activity of *Plodia interpunctella* (Lepidoptera: Pyralidae) in and around Flour Mills. *Journal of Economic Entomology*, **93**, 1842-1847. http://dx.doi.org/10.1603/0022-0493-93.6.1842
- [10] Johnson, J.A., Wang, S. and Tang, J. (2003) Thermal Death Kinetics of Fifth-Instar *Plodia interpunctella* (Lepidoptera: Pyralidae). *Journal of Economic Entomology*, **96**, 519-524. <a href="http://dx.doi.org/10.1093/jee/96.2.519">http://dx.doi.org/10.1093/jee/96.2.519</a>
- [11] Campbell, J.F. and Mullen, M.A. (2004) Distribution and Dispersal Behavior of *Trogoderma variabile* and *Plodia interpunctella* outside a Food Processing Plant. *Journal of Economic Entomology*, 97, 1455-1464. http://dx.doi.org/10.1093/jee/97.4.1455
- [12] Lord, J.C. (2001) Desiccant Dusts Synergize the Effect of *Beauveria bassiana* (Hyphomycetes: Moniliales) on Stored-Grain Beetles. *Journal of Economic Entomology*, **94**, 367-372. <a href="http://dx.doi.org/10.1603/0022-0493-94.2.367">http://dx.doi.org/10.1603/0022-0493-94.2.367</a>
- [13] Sabbour, M.M., Abd-El-Aziz, S., El-Sayed, A. and Sherief, M. (2012) Efficacy of Three Entomopathogenic Fungi Alone or in Combination with Diatomaceous Earth Modifications for the Control of Three Pyralid Moths in Stored Grains. *Journal of Plant Protection Research*, **52**, 359-364. <a href="http://dx.doi.org/10.2478/v10045-012-0059-7">http://dx.doi.org/10.2478/v10045-012-0059-7</a>
- [14] St. Leger, R.J. (1993) Biology and Mechanisms of Insect-Cuticle Invasion by Deuteromycete Fungal Pathogens. In: Beckage, N.E., Thompson, S.N. and Federici, B.A., Eds., *Parasites and Pathogens of Insects*, Volume 2, Academic Press Inc., New York, 211-229.
- [15] Gupta, S.C., Leathers, T.D., El-Sayed, G.N. and Ignoffo, C.M. (1994) Relationships among Enzyme Activities and Virulence Parameters in *Beauveria bassiana* Infections of *Galleria mellonella* and *Trichoplusia ni. Journal of Inverte-brate Pathology*, 64, 13-17. http://dx.doi.org/10.1006/jipa.1994.1062
- [16] Butt, T.M. and Goettel, M.S. (2000) Bioassays of Entomogenous Fungi. In: Navon, A. and Ascher, K.R.S., Eds., Bioassays of Entomopathogenic Microbes and Nematodes, CAB International, Wallingford, UK, 141-195. http://dx.doi.org/10.1079/9780851994222.0141
- [17] Mewis, I. and Ulrichs, C. (2001) Action of Amorphous Diatomaceous Earth against Different Stages of the Stored Product Pests *Tribolium confusum*, *Tenebrio molitor*, *Sitphilus granarius* and *Plodia interpunctella*. *Journal of Stored Products Research*, 37, 153-164. <a href="http://dx.doi.org/10.1016/S0022-474X(00)00016-3">http://dx.doi.org/10.1016/S0022-474X(00)00016-3</a>
- [18] Akbar, W., Lord, J.C., Nechols, J.R. and Howard, R.W. (2004) Diatomaceous Earth Increases the Efficacy of *Beauve-ria bassiana* against *Tribolium castaneum* Larvae and Increases Conidia Attachment. *Journal of Economic Entomology*, 97, 273-280. <a href="http://dx.doi.org/10.1093/jee/97.2.273">http://dx.doi.org/10.1093/jee/97.2.273</a>
- [19] Michalaki, M.P., Athanassiou, C.H., Kavallieratos, N.G., Batta, Y.A. and Balotis, G.N. (2006) Effectiveness of *Metarhizium anisopliae* (Metschnikoff) Sorokin Applied Alone or in Combination with Diatomaceous Earth against *Tribolium confusum* DuVal Larvae: Influence of Temperature, Relative Humidity and Type of Commodity. *Crop Protection*, 25, 418-425. http://dx.doi.org/10.1016/j.cropro.2005.07.003
- [20] Fields, P.G. and Korunic, Z. (2000) The Effect of Grain Moisture Content and Temperature on the Efficacy of Diatomaceous Earths from Different Geographical Locations against Stored Product Beetles. *Journal of Stored Products Research*, 36, 1-13. <a href="http://dx.doi.org/10.1016/S0022-474X(99)00021-1">http://dx.doi.org/10.1016/S0022-474X(99)00021-1</a>
- [21] Athanassiou, C.G. (2006) Influence of Instar and Commodity in Insecticidal Effects of Two Diatomaceous Earth Formulations against Larvae of *Ephestia kuehniella* (Lepidoptera: Pyralidae). *Journal of Economic Entomology*, 99, 1905-1911. http://dx.doi.org/10.1093/jee/99.5.1905
- [22] Vardeman, E.A., Arthur, F.H., Nechols, J.R. and Campbell, J.F. (2006) Effect of Temperature, Exposure Interval, and Depth of Diatomaceous Earth Treatment on Distribution, Mortality, and Progeny Production of Lesser Grain Borer (Coleoptera: Bostrichidae) in Stored Wheat. *Journal of Economic Entomology*, 99, 1017-1024. <a href="http://dx.doi.org/10.1093/jee/99.3.1017">http://dx.doi.org/10.1093/jee/99.3.1017</a>
- [23] Korunic, Z. (1998) Diatomaceous Earth: A Group of Natural Insecticides. *Journal of Stored Products Research*, 34, 87-97. http://dx.doi.org/10.1016/S0022-474X(97)00039-8
- [24] Subramanyam, B. and Roesli, R. (2000) Inert Dusts. In: Subramanyam, B. and Hagstrum, D.W., Eds., Alternatives to Pesticides in Stored-Product IPM, Kluwer Academic Publishers, Boston, 321-380. http://dx.doi.org/10.1007/978-1-4615-4353-4 12
- [25] Cox, P.D. and Wilkin, D.R. (1996) The Potential Use of Biological Control of Pests in Stored Grain. Research Review 36, Home-Grown Cereals Authority, London.

- [26] Sait, S.M., Begon, M., Thompson, D.J., Harvey, J.A. and Hails, R.S. (1997) Factors Affecting Host Selection in an Insect Host-Parasitoid Interactions. *Ecological Entomology*, 2, 225-230. http://dx.doi.org/10.1046/j.1365-2311.1997.t01-1-00051.x
- [27] Abbott, W.S. (1925) A Method of Computing the Effectiveness of an Insecticides. *Journal of Economic Entomology*, **18**, 265-267. http://dx.doi.org/10.1093/jee/18.2.265a
- [28] LeOra Software (2006) POLO-Plus 1.0 Probit and Logit Analysis. LeOra Software, Petaluma.
- [29] Buda, V. and Peciulyte, D. (2008) Pathogenicity of Four Fungal Species to Indian Meal Moth, *Plodia interpunctella* (Hubner) (Lepidoptera: Pyralidae). *Ekologija*, **54**, 265-270. <a href="http://dx.doi.org/10.2478/v10055-008-0040-y">http://dx.doi.org/10.2478/v10055-008-0040-y</a>
- [30] Hluchy, M. and Samsinakova, A. (1989) Comparative Study on the Susceptibility of Adult *Sitophilus granarius* (L.) (Coleoptera: Curculionidae) and Larval *Galleria mellonella* (L.) (Lepidoptera: Pyralidae) to the Entomogenous Fungus *Beauveria bassiana* (Bals.) Vuill. *Journal of Stored Products Research*, **25**, 61-64. http://dx.doi.org/10.1016/0022-474X(89)90011-8
- [31] Adane, K., Moore, D. and Archer, S.A. (1996) Preliminary Studies on the Use of *Beauveria bassiana* to Control *Sito-philus zeamais* (Coleoptera: Curculionidae) in the Laboratory. *Journal of Stored Products Research*, **32**, 105-113. http://dx.doi.org/10.1016/0022-474X(96)00009-4
- [32] Rice, W.C. and Cogburn, R.R. (1999) Activity of the Entomopathogenic Fungus *Beauveria bassiana* (Deuteromycota: Hyphomycetes) against Three Coleopteran Pests of Stored Grain. *Journal of Economic Entomology*, **92**, 691-694. http://dx.doi.org/10.1093/jee/92.3.691
- [33] Bourassa, C., Vincent, C., Lomer, C.J., Borgemeister, C. and Mauffette, Y. (2001) Effects of Entomopathogenic Hyphomycetes against the Larger Grain Borer, *Prostephanus truncatus* (Horn) (Coleoptera: Bostrichidae), and Its Predator, *Teretriosoma nigrescens* Lewis (Coleoptera: Histeridae). *Journal of Invertebrate Pathology*, 77, 75-77. <a href="http://dx.doi.org/10.1006/jipa.2000.4986">http://dx.doi.org/10.1006/jipa.2000.4986</a>
- [34] Lord, J.C. (2007) Desiccation Increases the Efficacy of *Beauveria bassiana* for Stored-Grain Pest Insect Control. *Journal of Stored Products Research*, **43**, 535-539. <a href="http://dx.doi.org/10.1016/j.jspr.2007.03.002">http://dx.doi.org/10.1016/j.jspr.2007.03.002</a>
- [35] Meikle, W.G., Cherry, A.J., Holst, N., Hounna, B. and Markham, R.H. (2001) The Effects of an Entomopathogenic Fungus, *Beauveria bassiana* (Balsamo) Vuillemin (Hyphomycetes), on *Prostephanus truncatus* (Horn) (Col.: Bostrichidae), *Sitophilus zeamais* Motschulsky (Col.: Curculionidae), and Grain Losses in Stored Maize in the Benin Republic. *Journal of Invertebrate Pathology*, 77, 198-205. http://dx.doi.org/10.1006/jipa.2001.5015
- [36] Padin, S.B., Dal-Bello, G.M. and Fabrizio, M. (2002) Grain Losses Caused by *Tribolium castaneum*, *Sitophilus oryzae* and *Acanthoscelides obtectus* in Stored Durum Wheat and Beans Treated with *Beauveria bassiana*. *Journal of Stored Products Research*, **38**, 69-74. http://dx.doi.org/10.1016/S0022-474X(00)00046-1
- [37] Ignoffo, C.M. and Garcia, C. (1992) Influence of Conidial Color on Inactivation of Several Entomogenous Fungi (Hyphomycetes) by Simulated Sunlight. *Environmental Entomology*, 21, 913-917. <a href="http://dx.doi.org/10.1093/ee/21.4.913">http://dx.doi.org/10.1093/ee/21.4.913</a>
- [38] Braga, G.L., Flint, S.D., Miller, C.D., Anderson, A.J. and Roberts, D.W. (2001) Variability in Response to UV-B among Species and Strains of *Metarhizium* Isolated from Sites at Latitudes from 61°N to 54°S. *Journal of Invertebrate Pathology*, **78**, 98-108. <a href="http://dx.doi.org/10.1006/jipa.2001.5048">http://dx.doi.org/10.1006/jipa.2001.5048</a>