# Levels of Maternal Serum Alpha-fetoprotein and Beta Human Chorionic Gonadotropin in HIV Seropositive Pregnant Women Attending Antenatal Care at Nnamdi Azikiwe University Teaching Hospital Nnewi, Nigeria 

C. H. Udeogu ${ }^{1 *}$, M. C. Ugwu ${ }^{1}$, J. Okwara ${ }^{1}$, A. C. Ohale ${ }^{2}$, O. C. Ike ${ }^{3}$, N. C. Amadi ${ }^{1}$,<br>O. Ugwu ${ }^{1}$ and F. C. Eguogwu ${ }^{1}$<br>${ }^{1}$ Department of Medical Laboratory Science, College of Health Sciences, Nnamdi Azikiwe University, P.M.B 5001Nnewi, Anambra State, Nigeria.<br>${ }^{2}$ Department of Medical Laboratory Science, PAMO University of Medical Sciences, Port-Harcourt,<br>Rivers State, Nigeria<br>${ }^{3}$ Department of Industrial Chemistry, Enugu State University of Science and Technology, P.M.B 01660, Agbani, Enugu State, Nigeria.

## Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

## Article Information

Received 20 July 2020


#### Abstract

HIV infection remains a worrisome pandemic especially in developing countries like Nigeria, with an increasing number of infected women becoming pregnant, with added risk of pregnancy complications such as intrauterine fetal death, neural tube defects, and vertical transmissions. Hence this study assessed the levels of maternal serum alpha-fetoprotein (MSAFP) and beta human chorionic gonadotropin ( $\beta$-hCG) in HIV seropositive pregnant women, and their implication


for maternal and fetal health. A total of 86 patients were recruited for the study from the Antenatal Clinic of Nnamdi Azikiwe University Teaching Hospital (NAUTH) Nnewi. Forty three were HIV seropositive (Test group), and forty three were HIV seronegative (Control group). Maternal serum alpha-fetoprotein and beta human chorionic gonadotropin levels were assayed by Enzyme Linked Immunosorbent Assay. The results showed that MSAFP levels ( $3.14 \pm 2.80 \mathrm{ng} / \mathrm{ml}$ ) were significantly lower ( $\mathrm{P}<.05$ ) in HIV infected pregnant women when compared to HIV uninfected pregnant women ( $4.75 \pm 3.68 \mathrm{ng} / \mathrm{ml}$ ). There was no significant difference in the levels of $\beta-\mathrm{hCG}$ $(217.88 \pm 63.85 \mathrm{mlU} / \mathrm{mL})$ in the test group ( $\mathrm{P}=0.629$ ) as compared to controls ( $223.75 \pm 47.71$ $\mathrm{mlU} / \mathrm{mL}$ ). In the test group, increased CD4 counts were significantly associated with increased MSAFP and $\beta$-hCG levels in a positive fashion ( $P<.05$ ). Increase in Gestational age was also significantly associated with increased $\beta$-hCG levels ( $\mathrm{P}<.05$ ).
The measurements of MSAFP and $\beta$-hCG levels may be used alongside CD4 count in the assessment of maternal and fetal wellbeing. Human Immunodeficiency Virus by reducing the levels of MSAFP in HIV infected pregnant women increases the likelihood of pregnancy complications such as intrauterine fetal death.

Keywords: Acquired immunodeficiency syndrome; alpha-fetoprotein; beta-human chorionic gonadotropin; HIV infections; pregnancy; Down syndrome.

## 1. INTRODUCTION

Human immunodeficiency virus infection/ Acquired immune deficiency syndrome (HIV/AIDS) is a disease of the human immune system caused by infection with HIV [1]. The virus is transmitted primarily through unprotected sexual intercourse, contaminated blood transfusions, hypodermic needles, and from mother to child during pregnancy, delivery, or breastfeeding [2]. It is a retrovirus that primarily infects components of the human immune system such as CD4 T-cells, macrophages and dendritic cells [3]. It directly and indirectly destroys CD4 Tcells. Basically, HIV causes AIDS by depleting CD4 T-cells. This weakens the immune system and allows for opportunistic infections [4].

Several publications have shown that maternal HIV infection and antiretroviral treatment (ART) modify the values of the biochemical markers used for second trimester screening for Down Syndrome (i.e. $\beta$-hCG, MSAFP, and unconjugated estriol) [5]. Unexplained levels of maternal serum AFP and $\beta$-HCG are associated with an increased risk of most pregnancy complications [6]. Beta-hCG has been implicated in modifying Kaposi sarcoma in HIV positive patients and in reducing HIV load in human lymphocytes and human choriocarcinoma cells. These anti-HIV effects of $\beta$-hCG may contribute to the limited maternal-fetal transmission of HIV infection ( $25 \%-35 \%$ ) without treatment [7]. Such inhibitory action of $\beta$-hCG may be present at varying levels throughout gestation based upon the circulating levels of $\beta$-hCG and its production by the placenta [8].

Human alpha-fetoprotein is a tumour associated fetal glycoprotein (oncofetal) that is associated with certain malignant neoplasms and with fetal defects during ontogenic growth and differentiation [9]. During pregnancy, AFP plays a role in the development of the embryo and fetus, and has been employed as a biomarker in fetal, prenatal, and pediatric disorders such as Beckwith-Wiedemann syndrome, Fanconi's anemia, and pancreatic blastomas [10,11]. Serum maternal AFP is first detectable in ( $\approx$ $5 \mu \mathrm{~g} / \mathrm{L}$ which is also equivalent to $5 \mathrm{ng} / \mathrm{mL}$ ) at about the $10^{\text {th }}$ week of gestation. The concentration increases about $15 \%$ per week to a peak of approximately $35 \mu \mathrm{~g} / \mathrm{L}$ [12]. The maternal serum concentration then subsequently declines slowly until term. After birth, the maternal serum AFP rapidly decreases to less than $2 \mu \mathrm{~g} / \mathrm{L}$ in an infant, serum AFP declines exponentially to reach adult concentrations by the $10^{\text {th }}$ month of life [13].

Beta-Human Chorionic Gonadotropin is a pregnancy associated immunodulating hormone, which has been recently shown invitro to suppress reverse transcriptase activity in chronically infected lymphocytes and monocytes, and to block viral transmission resulting from cellcell contact between virus carrying lymphocytes and placental trophoblasts [7]. Beta-hCG is synthesized in the syncytiotrophoblast cells of the placenta. Minute amounts are also made in the pituitary of men and non-pregnant women, and like many other pituitary hormones, it is secreted in a pulsatile fashion [14].

An increasing number of HIV-infected women are becoming pregnant [15]. Among women infected
with HIV, $80 \%$ to $90 \%$ of them are of reproductive age [16]. HIV infected pregnant women are at increased risk of most pregnancy complications such as intrauterine fetal death, harbouring babies with trisomy syndromes (involving neural tube defects), and transmitting this infection to their unborn babies [6]. In most sub Saharan African countries including Nigeria, HIV infection has become a leading medical complication of pregnancy and cause of maternal and neonatal morbidity and mortality [17]. It therefore becomes necessary to assess any possible role HIV has to play in the serum concentrations of AFP and $\beta$ hCGin HIV seropositive pregnant women.

## 2. MATERIALS AND METHODS

### 2.1 Study Design

A total of 86 pregnant women aged 18 to 50 years were conveniently recruited for this study, comprising of 43 individuals as test group (i.e. HIV infected pregnant women), and 43 as control (Apparently healthy pregnant women who were not diagnosed of any diseased condition as at the time of sample collection). The sample size was calculated from the formula: $N=\left[T^{2} \times P(1-P)\right] / M^{2}$; where $N$ is the sample size, $T$ is the confidence interval of $95 \%$ (1.96), $P$ is $6.0 \%$ and is the prevalence rate of HIV seropositive pregnant women in Nnewi Metropolis [18], M is the margin of error $(5 \%=0.05)$.

> Hence $\mathrm{N}=\left[1.96^{2} \times 0.06(1-0.06)\right] / 0.05^{2}$ Therefore $\mathrm{N}=86$.

### 2.2 Specimen Collection and Analytical Methods

Bio-data, and medical history - gestation age (weeks), Age (years), BMI, Parity, Systolic and Diastolic blood pressure, and CD4 count (cell $/ \mu \mathrm{L}$ ) were obtained from maternal health records file. BMI was calculated as weight (in kilograms) divided by the square of height (in metres). A single non-fasting blood sample was used for MSAFP and $\beta$-hCG analyses. About 5 mls of blood samples were drawn from the ante-cubital space into labeled plain containers, allowed to clot, and centrifuged at 5000 rpm for 5 minutes. The sera were carefully transferred into labeled plain containers, and stored frozen. The assay was done 3 weeks after collection.

### 2.3 Study Site

The study was carried out at the antenatal clinic of Prevention to mother-to-child transmission of

HIV clinic unit of Nnamdi Azikiwe University Teaching Hospital, Nnewi Nigeria. The analyses of MSAFP and $\beta$-hCG was done using ELISA method at the department of Chemical Pathology, NAUTH, Nnewi Nigeria.

### 2.3.1 Inclusion criteria

HIV infected pregnant women aged between 1850 years, as well as age-matched HIV uninfected pregnant women were recruited for this study.

### 2.3.2 Exclusion criteria

Pregnant women with hypertension, diabetes, hepatitis B or C co-infections, tuberculosis, and other chronic systemic infections were excluded from this study.

### 2.4 Statistical Analysis

The data from this study were subjected to statistical analysis using SPSS package, version 23 and presented as mean $\pm$ standard deviation. Student's t-test (independent t-test) was used to test difference in mean values. The results obtained were presented in tables for clarity. The $p$ - value $\leq 0.05$ was considered statistically significant. Correlation was performed using the Pearson's correlation.

## 3. RESULTS

Table 1 shows the gestational age, age, BMI, parity, blood pressure, CD4 count, AFP and $\beta$ hCG in HIV positive and negative pregnant women in the study. The results are expressed in Mean $\pm$ SD. There was no significant difference ( $P$ $>.05$ ) in the gestational age (in weeks), age (in years), parity, systolic blood pressure of HIV seropositive pregnant women when compared with HIV seronegative pregnant women. The diastolic blood pressure, AFP and $\beta$-hCG of HIV seropositive pregnant women was significantly lower ( $P<.05$ ) when compared with HIV seronegative pregnant women.

Table 2 shows that in HIV seropositive pregnant women, that there was a very weak positive relationship between Gestational age and AFP concentration, which was not significant $(P=$ 0.439 ). There was a very weak positive relationship between Gestational age and $\beta$ HCG concentration, which was significant ( $P=$ 0.014 ). There was also a weak positive relationship between CD4 count and AFP level,

Table 1. The levels of gestational age, age, BMI, parity, blood pressure, CD4 count, AFP and $\beta$ HCG in HIV positive and negative pregnant women (Mean $\pm$ SD)

| PARAMETER | HIV positive pregnant <br> women $(\mathbf{N}=\mathbf{4 3})$ | HIV negative pregnant <br> women $(\mathbf{N}=43)$ | t- value | Sig. |
| :--- | :--- | :--- | :--- | :--- |
| Gestational age <br> (weeks) | $26.02 \pm 7.77$ | $26.45 \pm 8.49$ | 0.245 | 0.807 |
| Age (years) | $30.77 \pm 3.85$ | $29.47 \pm 4.90$ | -1.368 | 0.175 |
| BMI $\left(\mathrm{kg} / \mathrm{m}^{2}\right)$ | $27.56 \pm 4.75$ | $27.40 \pm 4.42$ | -0.162 | 0.872 |
| Parity | $3.20 \pm 1.42$ | $2.53 \pm 1.32$ | -2.266 | 0.026 |
| Systolic BP $(\mathrm{mm} / \mathrm{Hg})$ | $106.27 \pm 10.43$ | $109.81 \pm 12.09$ | 1.454 | 0.150 |
| Diastolic BP $(\mathrm{mm} / \mathrm{Hg})$ | $65.84 \pm 8.30$ | $71.02 \pm 9.63$ | 2.672 | $0.010^{*}$ |
| CD4 (cells $/ \mu \mathrm{L})$ | $559.18 \pm 285.50$ |  |  |  |
| AFP $(\mathrm{ng} / \mathrm{ml})$ | $3.14 \pm 2.80$ | $4.75 \pm 3.69$ | -2.30 | $0.024^{*}$ |
| $\beta-\mathrm{hCG}(\mathrm{mIU} / \mathrm{mL})$ | $217.88 \pm 63.85$ | $223.75 \pm 7.71$ | -0.49 | 0.629 |
| N: Number of subjects, BMI: Body mass index, BP: Blood pressure, CD4: Cluster of differentiation 4, AFP: A/pha |  |  |  |  |

N: Number of subjects, BMI: Body mass index, BP: Blood pressure, CD4: Cluster of differentiation 4, AFP: Alpha fetoprotein, $\beta$-hCG: Beta human chorionic gonadotropin. $P \leq .05$ : *significant differences between groups (Sig.)

Table 2. Correlation between Gestation age, CD4, BMI with AFP and $\beta$-HCG

| Variables | Study $(\mathbf{N}=\mathbf{4 3})$ |  | Control $(\mathbf{N}=\mathbf{4 3})$ |  |
| :--- | :--- | :--- | :--- | :--- |
|  | $\mathbf{r}$ | Sig. | r | Sig. |
| Gestation age vs. AFP | 0.120 | 0.439 | 0.196 | 0.208 |
| Gestation age vs. $\beta$-hCG | 0.368 | $0.014^{*}$ | -0.039 | 0.805 |
| CD4 vs. AFP | 0.349 | $0.047^{*}$ |  |  |
| CD4 vs. $\beta$-hCG | 0.415 | $0.016^{*}$ |  |  |
| BMI vs. AFP | -0.036 | 0.814 | 0.122 | 0.435 |
| BMI vs. $\beta$-hCG | 0.090 | 0.562 | 0.137 | 0.383 |

N: Number of subjects, BMI: Body mass index, BP: Blood pressure, CD4: Cluster of differentiation 4, AFP: Alpha fetoprotein, $\beta$-hCG: Beta human chorionic gonadotropin. $P \leq .05$ : *statistically significant (Sig.), r: Pearson's correlation coefficient
which was statistically significant ( $P=0.047$ ). Furthermore, there was a weak positive relationship between CD4 count and $\beta$-hCG concentration, which was statistically significant ( $P=0.016$ ). There was a very weak negative relationship between BMI and AFP, which was not significant $(P=0.814)$. Also, there was a very weak negative relationship between BMI and $\beta$-hCG, which was not significant $(P=0.090)$. However, in HIV seronegative pregnant women, there was a very weak positive relationship between gestational age and AFP levels, which was not significant ( $P$ $=0.208)$. There was also a very weak negative relationship between gestational age and $\beta$-hCG levels, which was not significant ( $P=$ $0.805)$. There was a very weak positive relationship between BMI and AFP, which was not significant ( $P=0.435$ ). Also, there was a very weak positive relationship between

BMI and $\beta$-hCG, which was not significant $(P=$ 0.383).

## 4. DISCUSSION

Human immunodeficiency virus is a retrovirus that infects and depletes components of the human immune system such as CD4 T-cells and macrophages, hence predisposing the infected individual to opportunistic infections [4]. Maternal HIV infection modify the values of the biochemical markers used for second trimester screening for Down Syndrome (i.e. $\beta$-hCG, AFP, and unconjugated estriol) [5]. Chandra et al. [6] reported that unexplained levels of maternal serum AFP and $\beta$-hCG are associated with an increased risk of most pregnancy complications. Gross et al. [19] reported elevations in maternal serum AFP levels in HIV infected pregnant women as compared to controls. Ukibe et al. [20]
demonstrated that there was no significant difference between MSAFP in HIV infected pregnant women as compared to controls. Spencer [21] reported no elevations in the maternal serum levels of both AFP and $\beta$-hCG.

In the present study, the mean MSAFP level in HIV infected pregnant women was significantly lower than that of the control group, thus contradicts the reports by Gross et al. [19] and Ukibe et al. [20]. Since normal circulating levels of maternal AFP is an indication of good fetal health [5], the decrease in MSAFP levels among HIV infected pregnant women as observed in this study, may indicate that these women are at increased risk of pregnancy complications [6], such as intrauterine fetal death. Although Le Meaux et al. [22] reported that HIV infected pregnant women had lower median MSAFP levels than did control women, but with no significant difference, this study reported a statistically significant difference between the mean MSAFP levels in the test and control groups.

Although Yudin et al. [5] showed that $\beta$-hCG multiples of the median (MoM) were significantly higher in HIV infected pregnant women as compared to uninfected women, Spencer [21] and Le Meaux et al. [22] reported that $\beta$-hCG MoM levels did not differ between HIV infected pregnant women and the control subjects. The present study agrees with the findings of Spencer [21] and Le Meaux et al. [22], which showed that the difference between the mean serum levels of $\beta$-hCG in the test and control subjects was not statistically significant. This may be because of the significant role which $\beta$-hCG plays in the reduction of viral load in HIV infected pregnant women $[7,8]$, hence these women continue to secrete $\beta$-hCG as a way of preventing vertical transmission from mother to fetus.

Gross et al. [19] reported a negative correlation between maternal serum $\beta$-hCG level and CD4 count. The above finding contradicts our finding, as there was a moderately positive correlation between CD4 counts and $\beta$-hCG in HIV infected pregnant women, and this positive correlation was significant. There was also from this study a significant positive correlation between CD4 counts and MSAFP levels in HIV seropositive pregnant women. Hence the measurement of AFP and $\beta$-hCG can be used alongside CD4 count for the assessment of maternal and fetal wellbeing.

The systolic and diastolic blood pressure of both the test and control groups was within the normal range (120/80), although it was lower in the test group.

The mean CD4 status of the HIV-infected pregnant women was appreciably normal, suggesting a healthy immune system, perhaps due to antiretroviral therapy.

## 5. CONCLUSION

HIV infection caused a significant reduction in MSAFP levels of infected individuals as compared to the age and sex-matched controls. The difference in MSAFP levels so obtained may have resulted from the HIV infection and may indicate danger to both maternal and fetal health. Since normal circulating levels of MSAFP is an indication of good fetal health, it can be concluded from this study that HIV infection by causing decreased MSAFP levels in HIV seropositive pregnant women, may have a potential of posing increased risk of pregnancy complications such as intrauterine fetal death.

## CONSENT AND ETHICAL APPROVAL

The ethical approval for this research was obtained from the Human Research Ethics Committee of Nnamdi Azikiwe University Teaching Hospital, Nnewi, Anambra State. The procedures were explained to the subjects, and written informed consent obtained from each subject before specimen collection.

## ACKNOWLEDGEMENTS

Our sincere gratitude goes to the family of Udeogu, C. H, especially Mrs. Offor Chimsom N., Engr. Udeogu Jerry C., Engr. Udeogu Ifeanyi C., Pharm. Udeogu Sylvan C., Mrs. Udeogu Ogechi F., Dr. Odunze C. A., Barr. Abraham T. F and Miss Anowa Chinaza B and Lolo Chief M.N. Ugwu. You all were the driving forces behind the success of this research and publication of this manuscript.

## COMPETING INTERESTS

Authors have declared that no competing interests exist.

## REFERENCES

1. Ugwu MC, Okogun GRA, Okoye CF, Ekebor KL, Nwafia CJ, Nnona AE. Human
serum protein and C-reactive protein levels among HIV infected subjects in Uromi and its environs in Edo, Nigeria. International Journal of Basic, Applied and Innovative Research. 2016;5(3):74-80.
2. Markowitz. Environmental and Occupational Medicine (4 ${ }^{\text {th }}$ Ed.) Wolters Kluwer/Lippincott Williams \& Wilkins. Philadelphia. 2007;745.
3. Maddon PJ, Dalgleish AG, McDougal JS, Clapham PR, Weiss RA, Axel R. The T4 gene encodes the AIDS virus receptor and is expressed in the immune system and the brain. Cell. 1986;47:333-348.
4. Stevenson M. HIV-1 pathogenesis. Nature Medicine. 2003;9:853-860.
5. Yudin MH, Prosen TL, Landers DV. Multiple-marker screening in human immunodeficiency virus-positive pregnant women: screen positivity rates with the triple and quad screens. American Journal of Obstetrics and Gynecology. 2003;189:973-976.
6. Chandra S, Scott H, Dodds L, Watts C, Blight C, Van Den Hof M. Unexplained elevated maternal serum AFP and $\beta$ - hCG and the risk of adverse outcomes. American Journal of Obstetrics and Gynecology. 2003;189(3):775-781.
7. Bourinbaiar AS, Nagorny R, Tan $X$. Inhibitory effects of $\beta-\mathrm{hCG}$ on HIV-1 transmission from lymphocytes to trophoblasts. Federation of European Biochemical Societies Letter. 1992; 309:82-84.
8. Polliotti BM, Gnall S, Laughlin T, Miller R. Inhibitory effects of hCG preparation on HIV infection of human placenta in vitro. Placental Supplement. 2002;23:S102106.
9. Bei R, Mizjweski GJ. Alpha-fetoprotein is more than a hepatocellular cancer biomarker: from spontaneous immune response in cancer patients to the development of an AFP-based cancer vaccine. Current Molecular Medicine. 2011;11:564-581.
10. Mizjweski GJ. Physiology of alphafetoprotein as a biomarker of perinatal distress: relevance to adverse pregnancy outcome. Experimental Biology and Medicine (Maywood). 2007;232:993-1004.
11. Mizjweski GJ. Alpha-fetoprotein (AFP)derived peptides as epitopes for hepatoma immunotherapy: a commentary. Cancer Immunology and Immunotherapy. 2009;58:159-170.
12. Haddow JE, Palomaki GE, Knight GJ, Canick JA. Principles of prenatal screening for Downs Syndrome: combining multiple biochemical markers and maternal age. Foundation for Blood Research Handbook. 1998;11(20):20-38.
13. Wu JT, Roan Y, Knight JA. Serum AlphaFetoprotein levels in normal infants: their clinical and physiological significance. In: Mizejewski, G., Porter, L.H. eds. AlphaFetoprotein and Congenital Disorders. Orlando, Fla: Academic Press. 1985; 111-121.
14. Odell WD, Griffin J. Pulsatile secretion of Human Chorionic Gonadotropin in normal adults. The New England Journal of Medicine. 1987;317(27):1688-1691.
15. UNAIDS. Report on the global AIDS epidemic: $4^{\text {th }}$ global report. Geneva; UNAIDS; 2004.
16. Centres for Disease Control and Prevention. Epidemiology of HIV/AIDSUnited States, 1981-2005. Morbidity and Mortality Weekly Report. 2006;55:589:592.
17. Gregson S, Gamett GP, Anderson RM. Is HIV likely to become a leading cause of adult mortality in sub-Saharan Africal? Journal of Acqiured Immunodeficiency Syndrome. 1994;7:839-852.
18. Ikechebelu JI, Udigwe G, Ngozi J. Factors that influence the acceptance of HIV testing by pregnant women in Nnewi Nigeria. Tropical Journal of Medical Research. 2006;10(1):1-5.
19. Gross S, Castillo W, Crane M, Espinosa B, Carter S, Deveaux R, Salafa C. Maternal serum alpha-fetoprotein and human chorionic gonadotropin levels in women with human immunodeficiency virus. American Journal of Obstetrics and Gynecology. 2003;188:1052-1056.
20. Ukibe NR, Onyenekwe CC, Anojulu AA, Ukibe SN, Ikechukwu EC, Ehiaghe FA, Emeje PI, Ezugwu UA. Possible changes in maternal plasma cortisol, Adrenocorticotropic Hormones, Pregnancy associated Plasma Protein-A and Alphafetoprotein in HIV Pregnant women at NAUTH Nnewi, Nigeria. Journal of Bioanalysis and Biomedicine. 2017; 9:250-255.
21. Spencer K. First and second trimester markers of fetal aneuploidy in pregnant women with HIV infection. Fetal Diagnosis and Therapy. 2011;29:135-138.
22. Le Meaux JP, Tsatsaris V, Schmitz T, Fulla Y, Launay O, Goffinet F, Azria E. Maternal
biochemical serum screening for Downs Syndrome in Pregnancy with Human Immunodeficiency Virus Infection. National

Centre for AIDS/STD Control and
Prevention, China CDC. Obstetrics and Gynecology. 2008;112(2):223-230.
© 2021 Udeogu et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:
The peer review history for this paper can be accessed here:
http://www.sdiarticle4.com/review-history/58661

